(

15

20

25

30

534 Rec'd PT/PTO 25 SEP 2000

WO 99/48913

TITLE

"PEPTIDE TURN MIMETICS" FIELD OF THE INVENTION

THIS INVENTION relates to new compounds designed to be

peptide turn mimetics, and to new compounds useful for the synthesis of
peptide mimetics, especially turn mimetics. Peptide mimetics are used to
reproduce the important structural and functional elements contained in a
bio-active peptide sequence principally in order to develop novel
pharmaceuticals with increased binding affinity, selectivity, stability and/or
oral bioavailability compared to the bio-active peptide.

BACKGROUND OF THE INVENTION

Reverse turns (beta and gamma turns and beta buldges) are localised on the protein surface (Kuntz, 1972) and are of importance in protein interactions (Rose et al., 1985; Chalmers and Marshall, 1995) (and references contained therein). In addition reverse turns are important structures of peptide hormones and other biologically active peptides and cyclic peptides.(Giannis and Kolter, 1993; Olson et al., 1993; Kessler et al., 1995)

Peptide mimetics and peptide turn mimetics have as their object the replacement of a peptide sequence (a peptide turn) with a new compound which retains the elements essential for biological activity, thereby enabling or facilitating the development of novel pharmaceuticals devoid of the inherent problems of peptides - namely flexibility and poor pharmacodynamics. The essential elements for biological activity are thought to be the peptide sidechain groups (Farmer and Arièns, 1982: Ball and Alewood, 1990), therefore a peptide mimetic should include the side chain groups to have the best chance of retaining biological activity. A peptide mimetic may then take the form of a framework for displaying sidechain groups in an appropriate arrangement.

The majority of reverse turns are beta turns. The generally accepted definition of the beta turn is a sequence of four residues where the distance between the alpha carbons of residue (i) and residue (i+3).

. دود د د و دود د

(defined as d) is less than 7Å, and the central residues (i+1, i+2) are nonhelical.(Lewis et al., 1973) The general structure is shown below and includes the phi (ϕ) and psi (ψ) backbone dihedral angles that are used to describe the conformation of the peptide backbone. conversion of the beta turn to a beta turn mimetic is also shown - the A schematic peptide backbone is here replaced by an undefined framework.

$$R^{2}$$
 $Ca(i+1)$
 R^{3}
 $Ca(i+2)$
 $Ca(i+2)$
 $Ca(i+3)$

Mimetic Framework

 R^{4}
 $Ca(i+3)$

General structure of a hydrogen bonded The four backbone dihedral angles traditionally used B-turn. in turn classification are indicated, and also the position of the 7A upper distance cutoff for the definition of \$\beta-turns. d used for

A schematic representation of a beta turn mimetic - the peptide backbone has been replaced by an alternative chemical framework. represented here by a rectangle

10 The gamma turn is generally defined by the presence of a hydrogen bond between C=O (i) and N-H (i+2) to form a pseudo seven membered ring as illustrated below (Milner-White, 1988). equivalent hydrogen bond is present in a beta turn a pseudo ten membered ring is formed.

15

General structure of a γ -turn, defined by the presence of a hydrogen bond between the C=O of the (i) residue and the N-H of the (i+2) residue, as indicated.

10

15

20

25

30

(

The chemical synthesis of a framework having four independent chiral groups each with a wide range of possible functionality (for example, a beta turn mimetic) is a very significant synthetic challenge (Nakanishi and Kahn, 1996) as illustrated by the the fact that most proposed beta turn mimetics either do not provide for the incorporation of any sidechain functionality, or provide for a limited range of functionality, and at a limited number of positions. Reference may be made to reviews by Ball and by Hölzemann for illustration of these points (Ball and Alewood, 1990; Hölzemann, 1991; Hölzemann, 1991). In the case of mimetics that do provide for the incorporation of sidechain functionality, the syntheses are often complex and lengthy, and most seriously may require a different synthetic method for different sidechain sequences (i.e. the synthetic method is not generic). For example, in the work of Callahan, Huffman and Newlander on gamma turn mimetics the synthetic method varied depending on the sidechain sequence required - a 10 step sequence for a Gly-Phe-Leu mimetic, 13 steps for Phe-Gly-Val and 21 steps for Ala-Phe-Ala (Huffman et al., 1988; Callahan et al., 1992; Newlander et al., 1993). Given that the possible combinations of three residue sequences of the 20 natural amino acids is 8000 (20x20x20), and 160,000 for the four residue beta turn sequence, such non-generic methods are of limited use. The methods of Callahan and Huffman were further hampered by a lack of chiral control, as are most methods in the art.

In the development of peptide turn mimetics a further important issue is the reproduction of the variety of different turn conformations, particularly of the beta turn. Several different methods of describing turn conformation have been proposed, the traditional method having several turn types based on the backbone dihedral angles of the (i+1) and (i+2) residues i.e. I, I', II, II', III, III', IV, V, VIa, VIb, VII and VIII, with even this diversity of types being insufficient to adequately describe turn conformations.(Richardson, 1981; Wilmont and Thornton, 1990; Ball-

10

15

20

25

30

et al., 1993) No single mimetic framework can accurately mimic this diversity of turns; a selection of mimetic frameworks is required.

The problems encountered in the development of peptide turn mimetic syntheses are discussed in a review by Kahn (Kahn, 1993) and reference may also be made to a review article entitled "Design of Peptidomimetics" (Nakanishi and Kahn, 1996) which discusses aspects of mimetic design and developments regarding peptide mimetics.

The uses of reverse turn mimetics (and peptides or other compounds containing reverse turn mimetics) in drug development have been described in the art, notably in publications by Kahn and co-workers (Kahn, 1996; Nakanishi and Kahn, 1996; Qabar et al.,1996) and references contained therein. An important example of the application of reverse turn mimetics is the production of mimetics of known biologically active cyclic peptides (typically penta- or hexapeptides), as illustrated by Hirschmann and co-workers with O-D-glucose mimetics.(Hirschmann et al., 1992; Hirschmann et al., 1993) based

Other beta turn mimetics having biological activity are known in the art. For example, U.S. Patent 4535169 discloses a method for the synthesis of beta turn mimetics which can incorporate a functional substitution for the (i+3) sidechain (only), and Krystenansky et al. disclose a leucine enkephalin mimetic based on this method which had analgesic activity one third the potency of morphine (Krstenansky et al., 1982).

Reference may also be made to U.S. Patents 5475085 and 5618914 and International Publication WO96/22304 (all Kahn, M) which describe methods for the synthesis of a range of reverse turn mimetics. These mimetics are all produced by a modular synthesis technique (that may be applied to solid phase synthesis) which involves amino acid derivatives and various dipeptide azetidinones synthesised by a variety of techniques. An important common step in all of the syntheses of these mimetics is the cyclisation reaction which involves the azetidinone as activated ester component. Conformational variation is introduced to these mimetics by the inclusion of a variable component ("X") in the ring

10

15

20

25

30

of the cyclic turn mimetics. It should be noted that with two exceptions (the parent mimetics which have X=NH and have a ten or eleven membered ring) the beta turn mimetics produced by these methods have ring sizes of twelve members and above. Such large rings allow many conformations with d>7Å, the mimetic conformations are therefore biased away from the accepted definition of a beta turn (d less than 7Å), or more importantly the conformations are biased away from the most common reverse turn conformations which have d in the range of 4.5Å to 6Å (Rose et al., 1985; Gardner et al., 1993). Enkephalin mimetics have been made (Gardner et al., 1993) and also mimetics of a loop of CD4 that inhibit binding of HIV gp120 and infection of human lymphocytes (Chen et al., The synthetic methods described for the majority of these mimetics appear to be limited with respect to the possible functionality at the (i) and (i+1) positions, and indeed no mimetic with any functionality at the (i+1) position (other than -H = glycine = no sidechain) appears to have been described at this time.

Reference may also be made to International Publication-WO97/15577 (Kahn, M) which describes the synthesis of bicyclic reverse turn mimetics and chemical libraries containing such reverse turn mimetics. While concise, the synthetic methods do not provide for control of chirality at all positions, and the degree of sidechain function generality is questionable at two of the four positions. Furthermore the structure of the mimetics means they are not able to be easily incorporated in a peptide sequence, nor do they reproduce the relative positioning of the sidechain groups in the ideal manner (each sidechain attachment position should ideally be separated by three covalent bonds, as in a peptide).

Reference may also be made to the turn mimetics of Virgilio et al. (Valle et al., 1989; Virgilio and Ellman, 1994; Virgilio et al., 1996) that incorporate functionality at the (i+1), (i+2) and (i+3) positions (but not the (i) position), and that do not allow for incorporation of the mimetic in a peptide sequence (i.e. no amino and carboxy terminal groups in addition to the sidechains are present).

10

15

20

25

30

Reference may be made to U.S. Patents 5438188 and 5470849 (Callahan and Huffman) that describe biologically active compounds containing gamma turn mimetics, providing further illustration of the general utility of reverse turn mimetics.

Reference may also be made to International Publication WO95/25120 that describes the use of turn mimetics in the synthesis of peptide vaccines for generating a protective immune response in warm blooded animals.

In the methods and mimetics of the aforementioned references several common problems are evident: limited numbers of sidechains are able to be reproduced, there is limited control of chirality in the syntheses and a limited range of sidechain functions could be included. In addition, many of the syntheses of turn mimetics described are relatively long and complex, even when not all the sidechain functions are included, for example the syntheses of certain enkephalin mimetics were in the range of approximately 15 to 21 steps (Gardner et al., 1993). There is therefore still a need in the art for peptide mimetics that can incorporate a wide range of sidechain functions in all positions, that can be readily synthesised with control of chirality, and that have a wide range of conformations corresponding to those found in native peptides.

OBJECT OF THE INVENTION

It is the object of the invention to provide novel compounds useful as, and useful for the synthesis of, conformationally constrained mimetics of biologically active peptides and proteins (peptide mimetics). In particular, the invention provides new compounds and methods for the synthesis of new peptide reverse turn mimetics that can display a wide range of sidechain functions at all sidechain positions, can be incorporated in a peptide sequence, can be readily synthesised, and have a variety of conformations.

SUMMARY OF THE INVENTION

This invention describes novel compounds useful for the synthesis of peptide mirnetics, and describes the use of these compounds

10

15

20

25

for the synthesis of novel reverse turn mimetics. The reverse turn mimetics of the invention have the general structure X, or in a preferred embodiment the general structures I-VI (which are subsets of the general structure X; see below and Figures 1 and 2 on the attached sheets; the structures are fully described in the detailed description following this summary).

$$Q^2$$
 Q^3
 Q^1
 Q^4
 Q^3
 Q^1
 Q^4
 Q^2
 Q^3
 Q^4
 Q^2
 Q^3
 Q^4
 Q^2
 Q^3
 Q^4
 Q^4

It has now been discovered that B-allyldialkylboranes (e.g. Rg1a-i, Figure 3) react with imines 3 (Scheme 1) to give the novel allyl amines 4a-d in good yield and with a very high degree of chemo- and stereoselectivity. This is surprising because in contrast to these good results, allylation with the related B-allyldialkoxyboranes (e.g. Rg1j, Figure 3) or allylcopper or allylzinc reagents gave inferior results with racemisation and reaction at other functional groups. The reaction of imines 3 to form compounds 4a-d and formation of the related compounds 5-8a-d (all of which are made from compounds 4a-d) forms the basis of the synthesis of all the compounds of the invention, and hence the invention. Thus the allyl amines 4a-d are suprisingly valuable intermediates for the synthesis of new peptide mimetics, particularly reverse turn mimetics, enabling the synthesis of the significant variety of new reverse turn mimetics of the invention (having the general structure X), by the variety of different pathways described herein. All the mimetic systems of the invention can be incorporated into peptide sequences (i.e. they include amino and carboxy termini in addition to the sidechain

10

15

20

25

30

functions), or if desired the amino and/or carboxy termini can be omitted from the mimetic.

As described above, there is a need for a wide range of different mimetics to better reproduce the wide range of conformations found in native reverse turns. The turn mimetics of the invention have a large variety of novel functionalised ring structures, each of these therefore having novel conformational characteristics. Furthermore, the structure and ring sizes of many of the turn mimetics make them well suited to the reproduction of the geometry of the more common native reverse turn conformations (those having <u>d</u> of 4.5Å to 6Å).

The synthetic methods described in this invention are generally superior to the prior art in terms of the capacity to include a wide range of side chain functions, in all the sidechain positions, without significant changes in the synthetic method; that is, the methods are more truly generic. In addition, the control of chirality in the synthesis of the mimetics of the invention is superior to the prior art - an important consideration in the elucidation of structure-activity relationships and the development of novel pharmaceuticals, and other commercially useful peptide mimetics, as diastereomeric mixtures are normally unsuitable and may be impractical or impossible to separate on a commercial basis. Furthermore, selective access to a range of different diastereomers for a particular mimetic with a given sequence provides a selection of different conformations. Thus in a mimetic with four chiral centres there are a total of 16 (24) possible diastereomers - each having a different conformation. The methods of the invention allow for a high level of chiral control by using available chiral starting materials, non-racemising conditions and diastereoselective reactions.

The invention includes all novel intermediates used in the preparation of the turn mimetics and more generally useful for the preparation of peptide mimetics, particularly 4-8(a-d), Scheme 1 and 10, Scheme 2. Also 11-12, Scheme 3; 13-14, Scheme 4; 16-17, Scheme 5; 18-19, Scheme 6; 21-22, Scheme 7; 23(a-d)-25(a-d), 26, Scheme 8; 27-

15

20

25

28, Scheme 11; 29-34, Scheme 12; 35(a-c), 36-38, Scheme 13; 43-46, Scheme 15.

DETAILED DESCRIPTION OF THE INVENTION

The peptide mimetics of this invention have the general structure **X**, shown below and defined as follows:-

wherein R and R² and other R groups referred to hereinafter inclusive of R^1 , R^3 , R^4 , R^{n+3} and R^{n+4} etc. unless otherwise indicated, are amino acid side chain groups, each independently chosen and therefore the same or different (two separate R groups in the same mimetic do not require a different suffix to indicate that they are independently chosen and can be the same or different). The definition of "amino acid side chain group" as used in this document is the same as the definition of "amino acid side chain moiety or derivative" as described in International Publication WO97/15577, pages 7-9 (Kahn, M), incorporated herein by reference. Amino acid side chain groups typically correspond to, but are not limited to, those found in natural amino acids and derivatives and in common unnatural amino acids. Thus for glycine R = hydrogen; for for phenylalanine $R = -CH_2Ph$; for alanine R = methyl; homophenylalanine $R = -CH_2CH_2Ph$; for valine $R = -CH(CH_3)_2$; leucine $R = -CH_2CH(CH_3)_2$; p-nitrophenylalanine $R = -CH_2((4-NO_2)Ph)$; naphthylalanine R = -CH2-naphthyl etc. Also included are cyclic amino acid sidechains such as for proline, hydroxyproline and homoproline which involve a cyclization to the adjcent backbone nitrogen atom or the equivalent position, but only where this is possible (i.e. the amine or

15

20

25

equivalent atom is not already substituted as part of the heterocyclic mimetic framework).

Z is normally hydrogen, methyl, ethyl, formyl or acetyl, and may alternatively be R or - CH_2R or -C(O)R where R is an amino acid side chain group, or alternatively Z is part of a cyclic amino acid side chain group joined to R^2 (for example to mimic a proline residue at position (i+1)). For II(i) referred to hereinafter, Z cannot be hydrogen due to compound instability.

R^C is the carboxy terminal part of the mimetic, typically - 10 C(O)Pg^C or alternatively hydrogen or an amino acid side chain group R or -CH₂R.

Pg^C (and Pg^{C'} etc.) is a protecting group for carboxylic acid, typically including, but not limited to: alkoxy, benzyloxy, allyloxy, fluorenyl methyloxy, amines forming easily removable amides, or alternatively an appropriate cleavable linker to a solid phase support, or such a support itself, or alternatively hydroxy –OR, -NHR or remaining C-terminal portion of the mimetic system as described below.

 R^N is the amino terminal part of the mimetic, i.e. $-N(Z')Pg^N$,

Z' is normally hydrogen, alternatively methyl (to mimic an N-methyl amino acid residue at position (i)), or alternatively part of a cyclic amino acid side chain group joined to R¹ (for example, to mimic a proline residue at position (i)).

PgN (and PgN) is a protecting group for amine, typically including, but not limited to: Boc, Cbz, Fmoc, Alloc, trityl; or alternatively an appropriate cleavable linker to a solid phase support, or such a support itself, or alternatively hydrogen or R or -C(O)R where R is an amino acid side chain group, or alternatively part or all of the remaining N-terminal portion of the mimetic system, as described below.

M', M" are normally hydrogen, alternatively one or more may be C₁-C₄ alkyl (preferred methyl), chloro, C₁-C₄ alkoxy (preferred methoxy).

10

15

20

 $Q^1=R^1 \text{ and } Q^2=Z; \text{ alternatively there is a cyclisation from } Q^1 \text{ to } Q^2 \text{ and then in preferred embodiments of the invention } Q^1Q^2=CH(R)C(O) \text{ or } -CH_2CH(R)C(O)-\text{ or } -CH_2CH(R)C(O)-\text{ } Q^1Q^2 \text{ can also } De: -CH(R)CH_2-\text{ or } -CH_2CH(R)CH_2-\text{ or } -CH_2CH(R)CH_2-\text{$

 Q^5 = hydrogen, C_1 - C_4 alkyl, chloro or C_1 - C_4 alkoxy and Q^3 = Y or -C(O)NHCH(R)Y- or -C(O)ENHCH(R)Y-; or alternatively when Q^3 = -C(O)N(Q⁵)CH(R)Y- Q^5 is a covalent bond from the Q^4 group to the nitrogen atom in Q^3 (a cyclisation-forming a bicyclic ring system).

Y is selected from the group consisting of C(O) and CH_2 and Q^4 is selected from the group consisting of CHM^1 , C(O), $CH(Q^5)CH_2$ and $CH(Q^5)C(O)$ with the provisos that:

- (i) $Q^4 = CH(M^1), Y is C(0);$
- (ii) $Q^4 = C(O), Y \text{ is } CH_2;$
- (iii) $Q^4 = CH(Q^5)CH_2$, Y is C(O); and
- (iv) $Q^4 = CH(Q^5)C(O)$, Y is CH_2 .

 $E=-(AA)_n$ - where n=1, 2, 3, 4... (n=1 to about 300, but more typically n is between 1 and 30) and AA is an amino acid residue (e.g. $AA=-NHCH(CH_3)C(0)$ - for alanine); E is therefore a loop of n amino acids which are linked in a cycle by the rest of the mimetic system. The loop may also incorporate non-alpha amino acids, alpha dialkyl amino acids or any other amino acid which confers favourable properties on the mimetic system, for example increased binding affinity, or ease of detection, identification or purification. The invention, when used with such larger loops, is functioning as a covalent hydrogen bond mimic (another aspect of the invention), as generally described by Arrhenius *et al.* (Arrhenius *et al.*, 1987) and also in U.S. Patent 5807979 (Arrhenius *et al.*).

25

Preferred embodiments of the invention are the structures I-VI, as ilustrated in Figures 1 and 2 and defined in Table 1:-

Table 1

10

15

Q ¹	Q ²	Q ³	.Q5
R ¹	Z	Y	-
R ¹	Z	-C(O)NHCH(R)Y-	M
=	=		
R ¹	Z	-	M
:		C(O)NHCH(R)C(O)-	
		NHCH(R)Y-	
R ¹	Z	-C(O)N(Q5)CH(R)Y-	Q ³
			·
-CH(R)C(O)Q ²	Q1	Υ Υ	M
-CH ₂ CH(R)C(O)Q ²	Q1	Y	M
	R1 R1 -CH(R)C(O)Q2	R1 Z R1 Z R1 Z R1 Z -CH(R)C(O)Q2 Q1	R1 Z -C(O)NHCH(R)Y- R1 Z -C(O)NHCH(R)Y- C(O)NHCH(R)C(O)- NHCH(R)Y- R1 Z -C(O)N(Q ⁵)CH(R)Y- -CH(R)C(O)Q ² Q ¹ Y

Recursive entries of Q groups in Table 2 indicate a cyclisation - thus mimetics V and VI have a cyclisation between Q^1 and Q^2 , and mimetic IV has a cyclisation between Q^3 and Q^5 . In the Tables, the groups Q^1 - Q^5 and Y are as defined above, and the other groups are asdefined herein.

The compounds of this invention have been designed to allow for incorporation in a peptide or protein chain, or for covalent attachment to any molecule or group that may be useful for the enhancement of the biological activity, or other property, of the peptide mimetic. Thus the mimetics typically contain amino and carboxy termini independent of the sidechain functions. The term "remaining C- (or N-)

10

15

20

25

(

terminal portion of the mimetic" is any group, molecule, linker, support, peptide, protein, nucleoside, glycoside or combination of these, covalently linked to the mimetic. Typically such remaining portions would be peptides or combinations of peptides and other mimetics, or compounds to facilitate detection or identification, or to improve the pharmacodynamics or other useful feature of the mimetic system.

In addition, any R group (an amino acid side chain group) may serve as an attachment point to a solid support, or to a linker to a solid support, or as a covalent attachment point for another molecule that may be useful for the enhancement of the biological activity, or other property, of the mimetic, as described above for the remaining C- or N-terminal portions of the mimetic.

The term "cleavable linker" and "solid phase support" are as defined in International Publication WO97/1557

The use of a wavy line for one of the bonds at a chiral centre in the general structures X and I-VI and in the other structures in the Figures and Schemes indicates that the centre may be in either the (R) of (S) configuration, or be a mixture in any proportion of the (R) and (S) configurations. In most circumstances it is preferable to avoid mixtures of configurations unless the intention is to provide a mixture of diastereomers for example for the purpose of more efficient screening (by the use of a mixture) or for synthetic expediency. Chirality at the amino acid side chain positions in the compounds of the invention (e.g. at R1 to R4) is controlled by the use of chiral starting materials (L or D amino acids) and the avoidance of synthetic conditions which cause racemisation. The configuration at chiral centres formed in the mimetic synthesis is dependent on several factors and can be controlled in several cases, but in other cases mixtures of diastereomers will result, which can potentially be separated by physical means. A significant advantage of the invention is the superior level of chiral control possible at the chiral centres in the mimetics.

30

10

EXAMPLES OF PREFERRED EMBODIMENTS OF THE MIMETICS

□-Turn mimetics I(i)a, I(ii)a (M, M', M'', Z and Z' = hydrogen):

□-Turn mimetics II(i)a, II(iii)a (M, M', M" and Z' = hydrogen, Z = Me):

□-Bulge mimetics III(i)a, III(iii)a (M, M', M" and Z' = hydrogen, Z = Me):

(

10

15

Bicyclic □-turn mimetics IV(i)a, IV(ii)a (M, M', M", Z and Z' = hydrogen):

$$R^{2}$$
 R^{3}
 R^{4}
 R^{1}
 R^{1}
 R^{1}
 R^{2}
 R^{3}
 R^{2}
 R^{4}
 R^{1}
 R^{1}
 R^{2}
 R^{4}
 R^{1}
 R^{2}
 R^{2}
 R^{4}
 R^{1}
 R^{2}
 R^{4}
 R^{4}
 R^{1}
 R^{2}
 R^{2}
 R^{4}
 R^{4

Bicyclic □-turn mimetics V(i)a, VI(i)a, V(ii)a, VI(ii)a (M, M'and M" = hydrogen)-

$$R^{2}$$
 R^{1}
 R^{1}
 R^{1}
 R^{2}
 R^{1}
 R^{2}
 R^{1}
 R^{2}
 R^{1}
 R^{2}
 R^{2}
 R^{2}
 R^{2}
 R^{3}
 R^{2}
 R^{2}
 R^{3}
 R^{2}
 R^{3}
 R^{2}
 R^{4}
 R^{1}
 R^{2}
 R^{1}
 R^{2}
 R^{2}
 R^{3}
 R^{2}
 R^{2}
 R^{3}
 R^{2}
 R^{4}
 R^{1}
 R^{2}
 R^{2}
 R^{3}
 R^{4}
 R^{1}
 R^{2}
 R^{1}
 R^{2}
 R^{2}
 R^{3}
 R^{4}
 R^{1}
 R^{2}
 R^{2}
 R^{3}
 R^{4}
 R^{1}
 R^{2}
 R^{3}
 R^{4}
 R^{4}
 R^{1}
 R^{2}
 R^{3}
 R^{4}
 R^{1}
 R^{2}
 R^{3}
 R^{4}
 R^{4}
 R^{4}
 R^{4}
 R^{4}
 R^{1}
 R^{2}
 R^{3}
 R^{4}
 R^{4

The synthesis of all the mimetics described in this specification may proceed initially by the same general synthetic procedure for formation of the common intermediates - reaction of imines 3 with allyl metal reagents Rg1 (allyl boranes preferred) to give the allyl diamines 4, which are new, as described in Scheme 1. The other compounds of Scheme 1 (i.e. 5-8) may all be derived from the allyldiamines 4, as described in Scheme 1 and in the comments below. The

-12-3 -2-3

5

10

15

20

allylation reaction of imines 3, which falls within the scope of the invention, is remarkable for its mildness and selectivity - allowing a wide range of functional groups to be present in the rest of the molecule, a very important consideration in the synthesis of peptide mimetics. Another important feature of the reaction of allylboranes with the imines 3 is that it proceeds in good yield (e.g. >50% isolated yield) in the sterically hindered general case where R¹ and R² are both not hydrogen - i.e. for all mimetics of dipeptides not containing glycine. Scheme 1 and all subsequent Schemes describe the preferred case of RN=NHPgN and RC=C(O)PgC (Figures 1 and 2), analogous methods apply in the general case.

In relation to Scheme 1, preparation of the imines 3 is completed by condensation of an amino acid aldehyde (compound 1) with an amine (2a-d). The aldehydes 1 may be prepared by either oxidative procedures from the corresponding N-protected amino alcohol, or reduction of an N-protected amino acid derivative (Fehrentz and Castro, 1983), the different approaches have been reviewed, (Jurczak and Golebiowski, 1989) (see also Goel et al., 1988, Org. Syn. 67 69). The amines 2a are amino acid esters (or other acid protected amino acid derivatives), which are commercially available or may be synthesised by standard procedures from amino acids. Amines 2b-2d are prepared by reductive amination of an amine 2a and an amino acid aldehyde 1:

10

15

20

(

Amines 2d are prepared by repeated coupling/deprotection steps (as in conversion of 2b to 2c), standard techniques of peptide synthesis.

The reductive amination procedure for the alkylation of amines by aldehydes is well established in the art. (See for example, Sasaki and Coy, 1987, Peptides 8 119), the preferred reagents are sodium cyanoborohydride (Borch et al., 1971; Hutchins and Natale, 1979; Gribble and Nutatits, 1985), or more preferred sodium triacetoxyborohydride in dichloroethane. (Abdel-Magid et al., 1996).

Methods for the formation of amide bonds (coupling) are well established in the art. For coupling at more hindered amines the use of certain reagents, for example those based on 1-hydroxy-7-azabenzotriazole (Ehrlich et al., 1993; Carpino et al., 1994), or the use of amino acid fluorides (Carpino et al., 1990; Wenschuh et al., 1994) is advantageous.

Protecting strategies for the synthesis of peptides and peptide mimetics are well established in the art, for example a five dimensional orthogonal strategy was used by Hirschmann and co-workers in the synthesis of a somatostatin mimetic.(Hirschmann et al., 1996) A more general reference work on protection/deprotection is the monograph by Greene and Wuts.(Greene and Wuts, 1991).

10

15

20

25

30

The example syntheses described in this document use solution phase chemistry. The mimetics may also be synthesised by analogous solid phase techniques, or by a combination of solid phase and solution phase techniques, or the mimetics may be incorporated in normal solid phase peptide synthesis in the same way as a standard protected amino acid derivative. A review by Früchtel and Jung (Früchtel and Jung, 1996) details the state of the art in solid phase organic synthesis (in 1996).

It will be clear to those skilled in the art that the mimetics of the invention, due to their generic methods of synthesis, are suited to the application combinatorial chemistry techniques (more specifically combinatorial organic synthesis) and certain associated identification and screening techniques. The application of combinatorial and associated technologies to drug discovery are well known in the art and have been reviewed, see for example papers by Gallop et al. and by Gordon et al., and references therein, incorporated herein by reference (Gallop et al., 1994; Gordon et al., 1994). Additionally, reference may be made to a review by Thompson and Ellman on the synthesis and application of small molecule libraries, and references therein, incorporated herein by reference. (Thompson and Ellman, 1996).

The imines 3 form rapidly at room temperature on mixing of the amine and aldehyde in an appropriate solvent, e.g. CH₂Cl₂ or diethyl ether, with liberation of water. The water is removed with a drying agent, e.g. dried MgSO₄, which is subsequently removed by filtration. The imines are then reacted with an allyl metal reagent (Rg1) to give, after work-up, compounds 4 (Scheme 1).

In relation to reagents Rg1: standard allyl organometals, such as allyl magnesium bromide, are unsuitable for reaction with imines 3 due to a lack of selectivity for the imine function over the carboxylic acid derived groups (esters, amides) also present in 3. Allyl copper and zinc reagents have been used in selective reactions with imines (Bocoum et al., 1991; Basile et al., 1994) but in the case of imines 3 these reagents.

10

15

20

25

(

result in extensive racemisation at the D-imine chiral centre, and attack esters present in the imine to a significant extent. While some of the desired target 4 may be produced by many allyl metal reagents on reaction with 3, the reaction product typically contains a mixture of four diastereomers and also by-products from reaction at the carboxylic acid derived groups (especially esters). In contrast to these results, reaction B-allyl-9allyl boranes, such as with 3 the imines of borabicyclo[3.3.1]nonane (allyl-9-BBN), Rg1a, gives excellent results and reasonable diastereoselectivity (>50% isolated yield based on crude aldehyde, and ~80:20 diastereoselectivity where R1 is not H).

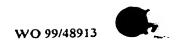
By the use of allyl trialkylboranes with appropriate chiral alkyl groups such as B-allyl-diisopinocampheylborane (allyl-DIP, Rg1b and Rg1c), or the diisocaranylboranes Rg1d-e it is possible to produce give only the major product (one diastereomer, >99:1) in good yield and purity. The configuration at the new stereocentre was determined to be (R) when using aldehyde derived from natural (S) configuration amino acids, and the stereocontrol exerted by the D-aldehyde chiral centre was dominant over the effect of chiral boron ligands and over the effect of the other amino acid chirality in all cases examined. The (+)DIP reagent Rg1b gave higher diastereoselectivity on imines derived from natural (S) The purity of the configuration aldehydes than Rg1c (from (-)DIP). allylation products 4a may also be improved by the removal of the ester protecting group Pg^C to give a crystalline amino acid which can be recrystallised (e.g. from ethanol/water) to the desired level of purity and then reprotected.

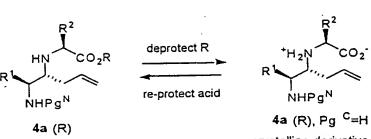
10

15

20

25





crystalline derivative The use of crotyl (Rg1f, Rg1h-i), methallyl (Rg1g) or other substituted allyl derivatives leads to bridge substituted mimetics (mimetics where at least one of M, M' and M" is not hydrogen) with further opportunities for stereocontrol. The less reactive allyl boronate allyldimethoxyboron (Rg1j) was found to give inferior results (significant epimerisation at C_(i) compared to the allyltrialkylboranes. allylboronate and related reagents (e.g. Rg1k-m) are described in the some of these may be more effective and literature, allyldimethoxyboron for the conversion of 3 to 4. Selective reactions using allylic metals have been reviewed by Yamamoto and Asao, Tables IV and V in the review (pp 2224-2230) list a wide variety of allyl boron reagents.(Yamamoto and Asao, 1993) The preparation of allyl-9-BBN and other allyltrialkylboranes has been described by Brown and coworkers (Kramer and Brown, 1977; Brown and Jadhav, 1983; Brown and Jadhav, 1984; Brown and Bhat, 1986; Brown, Randad et al., 1990) Allyltrialkylboranes may also be prepared by the reaction of the corresponding B-chloro or B-methoxy derivative with an allylmagnesium bromide (-78_C, diethyl ether), and reacted in situ with the imine (Yamamoto and Asao, 1993). The imines 3 formed from two non-glycine derivatives (i.e. R¹ and R² not H) are significantly hindered about the imine nitrogen, and the use of bulky boron ligands (such as diisopinocampheyl) can reduce the reaction yield. For high yield and selectivity smaller chiral B-allyl compounds, e.g. those based on 2,5dimethylboracyclopentane are preferred (e.g. Rg1n, Figure 3).

10

15

20

-25

In relation to protection and deprotection of compounds 4 and 5: addition of formaldehyde solution to 4 results in the rapid formation of imidazolidines 5; the relative configuration in the major allylation products 4 results in a 4,5-cis-substituted imidazolidine 5. This protection strategy is important for further reaction of these compounds. The protecting group is removed by treatment with aqueous acid (e.g. aqueous methanolic acetic acid).

A similar protection system is the dibenzyltriazone group of Knapp and co-workers, (Knapp et al., 1992) the paper describes other deprotection conditions and is incorporated herein by reference. An alternative deprotection method involves the hydrogenation of the imidazolidine system to an amine N-methyl group (40psi H_2 , Pd-C, MeOH, 48hrs), a conversion that can be used to give mimetics where Z = Me.

In relation to oxidation of alkenes 5: acids 6 can be synthesised directly by oxidative cleavage of the alkenes 5, e.g. by RuCl₃/NalO₄; aldehydes/ketones 8 may be synthesised directly from 5 by ozonolysis (for oxidation methods see for example the monograph by Hudlicky (Hudlicky) and references therein), but this process is not sufficiently selective and results in over-oxidation and the formation of other by-products. Preferred is the two step process of dihydroxylation (OsO₄, N-methylmorpholine-N-oxide (NMO),fBuOH/water) (VanRheenen et al., 1976; Ray and Matteson, 1980) to 7 followed by oxidative cleavage (Pb(OAc)₄ in benzene or H₅IO₆ in THF).(Hudlicky, 1990) Examination of the products of the oxidation reactions led to the surprising discovery that cleavage with (Pb(OAc)₄ resulted in isomerised product with the 4,5-substituents now trans. not cis as in the starting material. It was further

10

15

20

25

discovered that oxidation of the diol with H_5IO_6 in dry THF resulted in retention of the 4,5-cis configuration in the aldehyde product 8. The cis aldehydes can also be isomerised to the trans by treatment with catalytic acid, e.g. HCl in CHCl₃.

These important discoveries now allow selective access to all of the eight possible diastereomers of the aldehydes 8 and the acids 6, and therefore control of the majority of the chirality in all the mimetic systems described in the invention.

In relation to the oxidation of aldehydes 8 to acids 6: many oxidation reagents may effect this conversion, e.g. pyridinium dichromate (Hudlicky, 1990) Glycols 7 may also be oxidised directly to acids, e.g. by RuCl₃/NalO₄. In relation to reduction of acids 6 to aldehydes 8: carboxylic acids 6 can be converted to aldehydes by the same general methods used for the formation of protected □-amino aldehydes described above (Jurczak and Golebiowski, 1989). The carboxylic acid can be selectively reduced to the alcohol in the presence of carboxylic esters by the use of borane (Brown and Krishnamurthy, 1979), then oxidised to the aldehyde as previously described (Jurczak and Golebiowski, 1989)

In relation to **Scheme 2**: Aldehydes/ketones **8** undergo reductive amination with amino esters **9** by the methods previously described. The preferred method is NaBH(OAc)₃ in dichloroethane (room temperature). Surprisingly, it was discovered that the reductive amination of 4,5-cis imidazolidine aldehydes **8** resulted in the formation of the 4,5-

10

15

(

trans amines 10 (~9:1 trans:cis). This isomerisation reaction is rapid (much faster than that of aldehydes 8) as the reductive amination reaction is complete in only a few minutes. It was further discovered that the isomerisation reaction could be prevented by the pre-formation of the imine between the aldehyde 8 and amine 9 (in MeOH, 2-4 h at room temperature) with rigorous exclusion of acid, followed by reduction with sodium borohydride to give the cis amine 10 from the cis aldehyde. This discovery allows the selective synthesis of either the 4,5-cis diastereomer or 4,5-trans (9:1 with cis) diastereomer of the amines 10 starting from the 4,5-cis aldehyde 8.

It is important to appreciate that the methods described above allow the selective synthesis of all sixteen relative and absolute diastereomers of compounds 8 and 6, and all thirty two diastereomers of compounds 10. The ability to selectively synthesise these diastereomers is a significant advantage of the invention.

In relation to Scheme 3: Deprotection of 10 is by standard
methods consistent with the overall protecting strategy, as previously discussed. Many coupling agents are suitable for effecting the cyclisation

: 5-

Ĺ

15000

5

10

15

20

25

30

of 11 to 12, typical conditions: THF, BOP or HBTU or HATU. $EtN(i-Pr)_2$ (DIEA). The imidazolidine group is then deprotected (as previously described) by hydrogenation (MeOH, H₂-Pd/C) when Z = Me, and by hydrolysis (H⁺, H₂O) for Z = H (other Z groups may be introduced by acylation or alkylation of the deprotected secondary amine).

In relation to Scheme 4: Deprotection and cyclisation of 6b to 13, 14 and I(ii): - standard deprotection and coupling (cyclisation) methods are used. Other conversions are as previously described.

In relation to **Scheme 5**: As previously discussed, coupling reactions to relatively hindered (usually secondary) amines often require the use of specialised coupling conditions such as acid fluorides **15**, as described by Carpino *et al.* (Carpino *et al.*, 1990; Wenschuh *et al.*, 1994) Protecting groups PgN and PgC (in **16**) are typically benzyloxycarbonyl (Cbz) and benzyl ester, simultaneously deprotected by hydrogenation (0.1M HCl in EtOH, H₂-Pd/C), cyclised using the BOP coupling reagent in THF or DMF, followed by conversion (deprotection) of the imidazolidine group to N-Me by hydrogenation as previously described.

In relation to **Scheme 6**: Standard deprotection/ coupling conditions as previously described.

In relation to Scheme 7: Where R⁴ is a D-branched amino acid side chain (such as in Valine) then the coupling of 6a and 20 may require the use of HATU or other system suitable for a hindered coupling when bulky sidechain groups are present, as previously discussed. Conditions and protecting groups for the conversion of 21 to 19 are the same as for the conversion of 16 to II(i), Scheme 5.

In relation to **Scheme 8**: Hydroboration of alkenes is well known in the art, see for example monographs by Brown (Brown, 1975; Pelter *et al.*, 1988) The resulting alkyl boranes can be oxidised to alcohols (using alkaline hydrogen peroxide, or in a preferred embodiment using trimethylamine oxide, or other amine oxide, to form the borate with subsequent liberation of the alcohol by transesterification) (Soderquist and Najafi, 1986). Alternatively, treatment of the borane with acid

10

15

20

(

dichromate or, in a preferred embodiment, with pyridinium chlorochromate (PCC) gives the aldehyde (Brown et al., 1980; Brown et al., 1986). The aldehydes so formed may be reductively aminated on to amines 9 by the methods previously described.

In relation to **Schemes 9-11**: Standard synthetic techniques, previously described.

Methods for the synthesis of beta bulge (n=1, III(i-iv)) and higher loop mimetics (n>1), follow the corresponding methods for the synthesis of beta turn mimetics II(i-iv). Appropriate protecting groups are chosen so that extra residues can be added to the system prior to cyclisation, as illustrated in Scheme 11 for the synthesis of a III(i) mimetic.

In relation to Scheme 12: Conversion of 1,2-diols 7 to epoxides 29 (dehydration) may be achieved with a number of reagents, for example triphenylphosphine and a dialkylazodicarboxylate (the Mitsunobu reagents) (Carlock and Mack, 1978; Robinson, Barry et al., 1983) or TsCl/NaOH/PhCH₂NEt₃+ Cl⁻.(Szeja 1985). The epoxides 29 alkylate amines 9 on warming in ethanol or DMSO solution to give the amino alcohols 30. The alcohol may then be oxidised to the ketone 32 by use of TPAP (tetrapropylammonium perruthenate) with Nmethylmorpholine-N-oxide in CH₂Cl₂/acetonitrile by the method of Griffith and Ley (Griffith and Ley ,1990). For 32 typically PgN'=Cbz and PgC'=Obenzyl, then by the use of catalytic hydrogenation conditions (EtOH, H₂-Pd/C) the protecting groups are both removed and intramolecular reductive amination of the free amine to the ketone occurs to give 33. Coupling using the BOP reagent (or other suitable conditions) followed by deprotection of the imidazolidine group as previously described gives the bicyclic mimetic IV(i). Alternative syntheses are possible with the use of mild oxidising reagents to convert the glycols to carbonyl compounds, followed by reductive amination (Frigerio and Sangostino, 1994).

In relation to **Scheme 13**: 1,2 diols can be oxidised without carbon-carbon bond cleavage by the use of certain mild reagents e.g. IBX ⁻⁻

30

25

فستويد

....

- 5

...

٠٠٠.

ĺ

5.

(Frigerio and Sangostino, 1994). Conversion of **35c** to **36** proceeds by intramolecular reductive amination, or alternatively **35a** can be reductively aminated onto **2b**, as indicated. Reductive amination, coupling and deprotection details are as previously described.

The syntheses for the bicyclic □-turn mimetic systems V and VI are accomplished from the corresponding □-turn mimetic systems I, where the R¹ side chain group is derived from an aspartic acid (VI) or glutamic acid (VI) derivative.

10

15

20

25

The synthesis of mimetics V and VI thus proceeds as in Scheme 1, with the aldehyde component 1 (Scheme 1) being of the form 1d or 1e (Scheme 14), with the R and Pg groups as previously defined. The synthesis follows the synthesis of \Box -turn mimetic systems I, and is completed by the method illustrated in Scheme 15.

In relation to the preparation of alkylated aspartic and glutamic acid derivatives 1d and 1e the alkylated derivatives 39-42 can be prepared by a number of methods known in the art. Selected methods are summarised in Schemes 16 and 17. Rapoport and co-workers have developed methods for the selective alkylation of N-phenylfluorenyl protected aspartic and glutamic acid derivatives (Koskinen and Rapoport, 1989; Wolf and Rapoport, 1989). A review by Sardina and Rapoport, and references contained therein, describe several methods for the synthesis of alkylated aspartic and glutamic acid derivatives, incorporated herein by

10

15

20

25

30

reference (Sardina and Rapoport. 1996). Derivatives **39-42** are converted to aldehydes **1d** and **1e** by the methods previously described for for the preparation of aldehydes **1**.

The use of standard chemical techniques, in particular the Arndt-Eistert homologation reaction (Meier and Zeller, 1975) and reductions of carboxylic acids to aldehydes (Jurczak and Golebiowski, 1989), and also the synthesis of ketones -C(O)R from amides -C(O)N(OMe)Me (Nahm and Weinreb, 1981), to modify the aspartic and glutamic acid or their alkylated derivatives, or the use of similar derivatives of non-natural amino-acids, such as homo-glutamic acid, enables the synthesis of the other compounds of the invention in which -Q1Q2- (in the general structure X) forms part of a cyclic system, defined -CH₂CH₂CH(R)C(O)- (from sidechain -Q1Q2as: homoglutamic acid); -CH(R)CH2- (from aspartic acid by reduction of the □-carboxylate and reductive amination); -CH₂CH(R)CH₂- (from glutamic acid by reduction of the \(\pi\)-carboxylate and reductive amination); CH₂CH₂CH(R)CH₂- (similarly from homoglutamic acid); (from an aspartic acid sidechain ketone -CH2C(O)R by reductive -CH2CH2CH(R)- (from a glutamic acid sidechain ketone amination); CH₂CH₂C(O)R by reductive amination); -CH(R)CH₂C(O)- (postalkylation sidechain homologated aspartic acid); -CH2CH(R)CH2C(O)-(post-alkylation sidechain homologated glutamic acid); -CH(R)CH2CH2or -CH₂CH(R)CH₂CH₂- (from reductive amination of reduced postalkylation sidechain homologated aspartic acid or glutamic acid derivatives).

In relation to **Scheme 18**: An alternative procedure for the synthesis of intermediate compounds **10** (or equivalent) can be used in the case where R¹ is hydrogen and M, M¹ and M¹ are also hydrogen, as described in Scheme 18. Compound **49** is available commercially with certain N-protecting groups or can be made by coupling N-protected glycine with N,O-dimethylhydroxylamine. Reaction with vinylmagnesium bromide in analogy to the general procedure of Rapoport and co-workers.

.

10

15

20

WO 99/48913

28

(Cupps et al., 1985; Boutin and Rapoport, 1986) results in formation of the □,□-unsaturated ketone 50. Conjugate addition of an amino acid ester 9 (0_C, THF) results in the formation of aminoketones 51 which can be N-protected by standard procedures to form ketones 52 before reductive amination of an amino acid ester 9 under the conditions described by Abdel-Magid et al. (Abdel-Magid et al., 1996) (NaBH(OAc)3, dichloroethane) to form 54. Deprotection to 55 and coupling gives the Dturn mimetics I(i)a (where R1=H) as indicated. Alternatively the aminoketones 51 can be acylated with an amino acid fluoride 15 to give compounds 53 which can be deprotected and cyclised (by reductive amination)-by hydrogenation in mild acid conditions (H2/Pd-C, 0.1M HCl in EtOH). The reductive amination-cyclisation is diastereoselective, only one diastereomer of the mimetics I(i)a were formed from 53, with the configuration at the new stereocentre controlled by the R2 stereocentre. The (S) configuration at R2 gives (S) at the new centre. In contrast, the 54 proceeds with lower reductive amination to form amines stereoselectivity (~3:1) with the major diastereomer having the (R) These discoveries provide further configuration when R2 is (S). opportunity for stereocontrol in the synthesis of the turm mimetics. Deprotection of compounds 54 and reaction with formalin in THF is an alternative method for synthesis of compounds 10 (R1=H), as described in Scheme 18.

10

dlpc₂Ballyl

Rg1b

EXAMPLE SYNTHESES

Synthesis of a D-turn mimetic I(i) by the general Example (A). procedure

mimetic for the sequence HTyr-Gly-Gly-Phe, which is found in the enkephalins, was synthesised with a D-turn mimetic based on the Tyr-Gly-Gly tripeptide. Similar mimetics have shown activity at opiate receptors (Huffman, Calishan et al., 1988; Huffman et al., 1989).

The synthesis is summarised in the following scheme:-

$$R = p-(OBn)Ph$$

$$R = p-(OBn)P$$

59

DIPEA,
EtOAc,
$$\Delta$$
Boc N
R
CO₂CH₃
Boc N
R
64
CO₂H

ĺ

(

4.2

5

10

15

20

25

30

Preparation of 56:

The amide 56 was synthesised from commercially avaliable Boc-Tyrosine(OBn)OH by coupling with N,O-dimethylhydroxylamine hydrochloride, 1 equivalent, in DMF/CH2Cl2 (1:5) using HBTU reagent (1 eq.) and DIEA (2 eq.) at room temperature. The CH2Cl2 was evaporated in vacuo and the residue partitioned between diethyl ether and aq. NaHCO3. The aqueous layer was separated and the ether layer washed in turn with 1M HCl (x2), aq. NaHCO₃, brine, and then dried over MgSO₄. Filtration and removal of the solvent in vacuo left the product amide 56 as a white crystalline solid in >90% yield. Further purification was carried out by silica gel chromatography eluting with ethyl acetate in petroleum ether, or by recrystallisation from ether. 1H NMR (300 MHz, CDCl₃): 🗇 7.46-7.28, 5H, m, OBn; 7.08, 2H, d, J=8.5 Hz, Tyr Ar; 6.90, 2H, d, J=8.5 Hz, Tyr Ar; 5.15, bd, J=8 Hz, NH; 5.04, 2H, s,)OCH₂Ph; 4.91, 1H, bm, Phe□; 3.65, 3H, s, OCH₃; 3.16, 3H, bs, NCH₃; 3.00, 1H, dd, J=6, 13.5 Hz, Phe□; 2.83, 1H, dd, J=7, 13.5 Hz, Phe□; 1.40, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃): □ 172.3; 157.6, Tyr Ar-O; 155.1, carbamate; 137.0_ ipso; 130.4; 128.8; 128.5; 127.8; 127.4; 114.7; 79.5, tBoc; 69.89, OCH₂Ph; 61.43, Tyr□; 51.55, OCH₃; 37.89, NCH₃; 32.00, Tyr□; 28.26, Boc.

Preparation of 57:

The aldehyde **57** was prepared by the method of Fehrentz and Castro (Fehrentz and Castro, 1983) as follows: to a stirred solution of 4.2 g of amide **56** in 100 mls of anhydrous diethylether cooled to 0°C was added 0.51 g lithium aluminium hydride. After 10 minutes a solution of 1.5g NaHSO₄ in 30 mls of water was added. The reaction mixture was diluted with more ether and washed with 1M HCI, saturated aqueous sodium bicarbonate and brine and dried over magnesium sulphate. The volatiles were removed under reduced pressure to give a waxy solid which was recrystallised from cold ether/hexane to give 2.6 g (72%) of **57** as a white solid. ¹H NMR (300 MHz, CDCl₃): □ 9.62, 1H, s, aldehyde; 7.50-7.25, 5H, m, Ar(OBn); 7.10, d, J=8 Hz, Ar(Tyr); 6.93, 2H, d, J=8 Hz,

10

15

20

25

30

(

Ar(Tyr); 5.10, 1H, b, NH; 5.05, 2H, s, OCH₂Ph; 4.39, 1H, q, J=7 Hz; Tyr□; 3.06, 2H, d(ABX), J=7 Hz, Tyr□; 1.44, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃): □ 199.6; 157.8, TyrOAr; 155.3, carbamate; 136.9, ipso; 130.3; 128.5, 127.9, 127.4; ArCH; 115.0, ArCHTyr; 80.08, tBoc; 69.69, OCH₂Ph; 60.82, Tyr□; 34.51, Tyr□; 28.22, Boc. Preparation of 58:

The imine 58 was formed by the reaction of the aldehyde 57 (1.4 g) with one equivalent of glycine benzyl ester in $10ml\ CH_2Cl_2$ (stir at room temperature 1 h) the water formed was removed with magnesium sulphate which was then removed by filtration.

1H NMR (300 MHz, CDCl₃): ☐ 7.68, 1H, s, imine; 7.49-7.30, 10H, Ar; 7.15, 2H, d, J=8 Hz, TyrAr; 6.92, 2H, d, J=8 Hz, TyrAr; 5.67, 1H, bd, J=6 Hz, NH; 5.20, 2H, s, OCH₂Ph; 5.05, 2H, s, OCH₂Ph; 4.51, 1H, bm, Tyr□; (4.26, 4.22), 2H, AB, J=15.5 Hz, Gly□; 3.15, 1H, bdd, J=5.0, 13.5 Hz, Tyrb; 2.93, 1H, dd, J=8.0, 13.5 Hz, Tyrb; 1.48, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃): ☐ 169.3; 167.4, CH imine; 157.5; 155.1; 136.9, 135.3: 2x ipso; 130.4, CHAr; 128.8, Tyr ipso; 128.44, 128.39, 128.26, 128.19, 127.76, 127.29, 114.65: ArCH; 79.22, tBoc; 69.81, TyrOCH₂Ph; 66.60, GlyOCH₂Ph; 60.48, Tyr□; 54.73, Gly□; 37.97, Tyr□; 28.23, Boc.

Preparation of 59:

(Rg1b) was prepared by the addition of allylmagnesium bromide to one equivalent of (+)DIP-CI in anhydrous diethyl ether under dry nitrogen. Brown and Jadhav, 1983). The solution of imine 58 in CH₂Cl₂ was stirred and cooled to -78°C under dry nitrogen and one equivalent of the previously prepared dlpc₂Ballyl solution added. The mixture was allowed to warm gradually to room temperature (overnight). The volatiles were removed under reduced pressure and the residue dissolved in THF and 1 ml of glacial acetic acid added. The mixture was refluxed overnight and then the volatiles removed under reduced pressure. The crude product was dissolved in CH₂Cl₂ / petroleum ether and the precipitate filtered off.

4.

(

2

5

10

20

25

30

The residual oil was chromatographed on flash silica eluting with ethyl acetate / petroleum ether to give 1.3 g (60% yield based on 57) of 59. TLC 1:2 EtOAc:light pet. Rf=0.40. 1H NMR (300 MHz, CDCl₃): 7.48-7.30: 10H, Ar; 7.13, 2H, d, J=8.5 Hz, TyrAr; 6.91, 2H, d, J=8.5 Hz, TyrAr; 5.84, 1H, m, vinyl CH; 5.17, 2H, s, TyrOCH₂Ph; 5.16, 2H, m, vinyl CH₂; 5.05, 2H, s, GlyOCH₂Ph; 4.90, 1H, bd, J=8.5 Hz, NHBoc; 3.95, 1H, bm, Tyr□; 3.54, 2H, s, Gly□; 3.82, 1H, dd, J=4.5, 14.4 Hz, Tyr□; 2.73, 3H, be; NH(amine), Tyr , CH(homoallyl); 2.28, 2H, m, allyl; 1.35, 9H, Boc. 13C NMR (75 MHz, CDCl₃): 0 172.1; 157.3; 155.6; 137.1, 135.4: ipso; 134.9, CHvinyl; 130.6, ipsoTyr; 130.0, 128.5, 128.4, 128.3, 127.8, 127.3: 114.7, TyrArCH; 79.05, tBoc; 69.90, 117.8, CH₂vinyl; ArCH: TyrOCH₂Ph; 66.51, GlyOCH₂Ph; 59.38, Tyr□; 53.46, CH; 49.28, Gly□; 35.44: coincident allyl carbon and Tyr□; 28.20, Boc. Mass Spectrum (ISMS) m/z 545.1 (MH+), calculated for C₃₂H₄₅N₃O₅: 544.

15 Preparation of 60:

The amine 59 (930 mg, 1.7 mmol) was dissolved in ethyl acetate (15 mL) and 37% aq. formaldehyde solution added (1 mL). The solution was stirred vigorously at room temperature for 1 h (or until the reaction was complete) and then diluted with ether (100 mL) and washed in turn with aq. NaHCO3, water (x3), brine and then dried (MgSO4). Removal of solvent in vacuo left an approximately quantitative yield (950 mg) of the crude product 60 which was used in the next reaction or further purified by flash chromatography eluting with 10-15% ethyl acetate in light petroleum. TLC 33%EtOAc:light pet. Rf=0.56. The NMR spectra were quite broad in CDCl3, amide rotamers were present in the approximate ratio 2:1. ¹H NMR (300 MHz, CDCl₃):

7.50-7.27, 10H, m's, Ar; 7.09, 2H, m, Ar; 6.90, 2H, d, J=8.5 Hz, Ar; 5.64, 1H, bm, vinyl CH; 5.19, 2H, s, OCH₂Bn; ~5.1, 2H, m, vinyl CH₂; 5.05, 2H, s, OCH₂Bn; 4.59, 1H, bm, ring NCH₂N(a); 4.17, 1H, bm, ring NCH₂N(b); 4.06, 1H, bm, Tyr ; 3.70, 1H, d, J=17 Hz, Gly□(a); 3.42, 1H, bd, J=17 Hz, Gly□(b); 3.16, 1H, bm, TyrC'H(ring); 2.84, 2H, bm, Tyr0; 2.31, 2H, m, allylCH₂; 1.38, ~3H, bs, Boc minor rotamer; 1.19, ~6H, s, Boc major rotamer. ¹³C NMR (75 MHz,

15

20

25

30

CDCl₃): $\Box\Box\Box$ (peaks due to the carbamate rotamers are placed in parentheses, major rotamer first) 169.8 (ester); 157.2 (tyrosine O-ipso); (153.1, 152.8) carbamate; 137.2 (ipso); 135.4 (ipso); 134.2 (CH vinyl); 131.3 (ipso); 130.5, 128.5, 128.4, 128.3, 127.8, 127.4, 127.3, 126.9: ArCH; 117.5 (vinyl CH₂); 114.7 (2xTyrArCH); 79.52 (Boc tertiary); 69.93 (CH₂); 66.95 (CH₂); 66.46 (CH₂); 64.27 (CH); (59.65, 58.76) (CH); 51.60 (CH₂); 34.34 (CH₂); (32.20, 31.93) (CH₂); (27.93, 28.25) (Boc 3xCH₃). Mass Spectrum (ISMS) m/z 557.1 (MH⁺), calculated for $C_{34}H_{40}N_2O_5$: 556 fragments (OR 60): 501.1, (-tBu).

10 Preparation of 61:

N-oxide (NMO), 40 mg of a 2.5% (by weight) solution of osmium tetroxide in *t*-butanol, 4 mls of *t*-butanol and 0.5 mls water. The mixture was stirred at room temperature until the reaction was complete (about 24 hours). 3 mls of 10% NaHSO₃ was added, the solution stirred for 10 minutes, then neutralised with sodium bicarbonate, diluted with brine and extracted three times with ethyl acetate. The combined extracts were washed with brine and dried over magnesium sulfate. Removal of volatiles under reduced pressure gave the crude diol in good yield as an oil which could be used in the next reaction or purified if required by chromatography on silica gel eluting with ethyl acetate. Mass Spectrum (ISMS) m/z 591.3 (MH+), calculated for C₃₄H₄₂N₂O₇: 590.

Oxidation of diol using Pb(OAc)₄: The diol (100 mg, 0.17 mmol) was dissolved in dry benzene (4 mL) and Pb(OAc)₄ (85 mg, moistened with acetic acid) was added. After 10 min stirring at room temperature the reaction was filtered, the solvent removed *in vacuo* and the residue purified by flash chromatography eluting with 25%EtOAc in light petroleum. Yield of the aldehyde 61 was 32% (30 mg). (No efforts to optimise the yield were made. Yield might be improved, for example, by partitioning the crude reaction mixture between aq.base and EtOAc to ensure none of the amine product was lost on filtration of the insoluble salts.) TLC 50%EtOAc in light pet. Rf=0.51. NMR analysis (NOESY)

5

10

20

25

30

experiment) indicated the 4,5-trans ring conformation (i.e. the 4(S) isomer). ¹H NMR (300 MHz, CDCl₃): \Box 9.52, 1H, t, J=1.5 Hz, aldehyde; 7.50-7.25, 10H, m, ArH; 6.92, 2H, d, J=9 Hz, TyrAr; 5.15, 2H, s, OCH₂Ph; 5.05, 2H, s, OCH₂Ph; 4.65, 1H, bm, ringCH₂(i); 3.88, 1H, bm, Tyr \Box ; 3.80, 1H, bm, ringCH₂(ii); 3.45, 1H, d, J=16 Hz, Gly \Box ; 3.44. 1H. m, ringCH(\Box aldehyde); 3.28, bd, J=16 Hz, Gly \Box ; 3.17, 1H, bm, Tyr \Box ; 2.80, 1H, dd, J=9.0, 13.5 Hz, Tyr \Box ; 2.51, 1H, J=6, 17 Hz, \Box aldehyde; 2.28, 1H, dd, J=17, 4.5 Hz, \Box aldehyde; 1.50, 9H, Boc. ¹³C NMR (75 MHz, CDCl₃), (rotamers): \Box 200.5; 169.9; 157.5; 153.1; 136.9; 135.3; 130.5, 129.6, 128.6, 128.5, 128.4, 127.6, 127.4, 115.0; Ar; 80.21, tBoc; 69.92, OCH₂Ph; (67.08, 66.86) br, CH₂; 66.58, OCH₂Ph; (62.93, 62.56) br, CH; (61.35, 60.72) br, CH; 52.14, CH₂; 46.36, CH₂; (38.5, 37.27) br, CH₂; 28.38, Boc. Mass Spectrum (ISMS) m/z 559.1 (MH+), calculated for C₃₃H₃₈N₂O₆: 558.

15 Preparation of 62 and 63:

The aldehyde 61 (30 mg, 500mol) was dissolved in 1.2dichloroethane (5 mL) and glycine methyl ester hydrochloride (50 mg) and NaBH(OAc)₃ (50 mg) added. The reaction was stirred at room temperature and was complete in a few minutes (<15 min). The reaction was diluted with ethyl acetate, and washed in turn with aq.NaHCO3, water, brine and then dried (MgSO₄). Evaporation of the solvent left the crude product 62 as a clear oil: TLC 1:1 EtOAc:light pet. Rf=0.17. Mass Spectrum (ISMS) m/z 632.3 (M+H $^+$), calculated for $C_{32}H_{45}N_3O_5$: 631 Analysis of the product or the reaction mixture after overnight standing revealed the formation of a new product with a mass spectrum corresponding to the target cyclised material 63 (MH+=524Da). Thus the amine product 62 was not generally isolated but converted directly to 63. The spontaneous cyclisation was accelerated by the addition of base (i-Pr₂NEt). After removal of solvent by evaporation under reduced pressure and the product was purified by flash chromatography eluting with 10-20% EtOAc in light pet. TLC: 1:1 EtOAc:light pet. Rf=0.51. 1H NMR (300 MHz, CD₃CN): [] 7.47-7.29, 5H, m, ArH; 7.12, 2H, m, Tyr; 6.92,

10

15

20

2H, m, Tyr; 5.07, 2H, s, OC \underline{H}_2 Ph; 4.35, 1H, d, J=5.4 Hz; ABq, □a=4.05, □b=4.02, JAB=17.4 Hz; 3.70-3.52, 6H, overlapped signals (includes: 3.65, 3H, s; 3.58, 1H, dd, J=11.2, 15.2 Hz); 3.49-3.32, 2H, br m's; 3.15, 1H, br dd, J=5.5, 15.5 Hz; 2.99, 1H, br dd, J=13.4, 14.9 Hz; 2.80, 1H, vbr m; 2.68, 1H, vbr m; 1.64, 1H, m; 1.46, 10H, s + m, Boc resonance obscures multiplet. ¹³C NMR (75 MHz, CD₃CN), rotamers, in approximate ratio 3:2, split some peaks and are recorded in parentheses: □ 173.3; 171.5; 158.8; 155.0, br; 138.9; 132.0; 129.9; 129.2; 129.0; 116.1; 80.84; 71.01; (70.87, 69.99); (68.12, 67.45); (65.47, 64.89); 55.76; 52.93; 51.45; 49.95; (39.00, 37.53); 31.87; 28.97 (Boc). Mass Spectrum (ISMS) m/z 524.3 (M+H+), calculated for C₂9H₃7N₃O₆: 523. Preparation of compounds 64 to 66:

The product 63 was hydrolysed with LiOH/H₂O/MeOH to the coupled then $MH^{+}=510$ and spectrum (mass acid 64 standard phenethylamine using (DMF/CH₂Cl₂/HBTU/DIEA) with procedures and work-up to give 65. The imidazolidine ring of 65 was deprotected with a solution of acetic acid-methanol-water (~1:1:1, stirred as a very dilute solution for several days then lyophilised) to give crude 66 as a white amorphous solid. Mass Spectrum (ISMS) m/z 601 (M+H+), calculated for C₃₅H₄₄N₄O₅: 600.

Example (B). Synthesis of a (4,5)-cis imidazolidine aldehyde by oxidation of a diol.

25

For the preparation of the 4,5-cis aldehyde 68 (in this case the 4(R) isomer) the diol 67 prepared from alkene 60 (as described above) (1mmol) was dissolved in THF (10 mL) and H_5IO_6 (1 mmol)

12.

5

10

15

dissolved in THF (~20 mL) was added and the reaction stirred at room temperature. A precipitate of iodic acid rapidly formed and the reaction was complete in <5 min. The THF solution was diluted with ether and washed in turn with 10% aq.Na₂CO₃, water, brine and then dried (MgSO₄). The product aldehyde 68 was formed in good yield and purity. Contact with acid should be minimised to prevent isomerisation to the trans aldehyde and/or decomposition, for example avoid chloroform as an NMR solvent unless recently made acid free. Yield was 60-80%. TLC: 50%EtOAc in light pet. Rf=~0.5. ¹H NMR (300 MHz, CD₃CN): ☐ (peaks moderately broad: the Boc rotamers were not resolved although the Boc peak was asymmetric and very broad) 9.48, 1H, bm, aldehyde; 7.5-7.3, 10H, m, 2xBn; 7.09, 2H, bd, J=7.5 Hz, Tyr Ar; 6.88, d, 8.2 Hz, Tyr Ar; 5.13, s, 2H, OCH₂Ph; 5.05, s, 2H, OCH₂Ph; 4.38, 1H, d, 6.0 Hz, NCH₂N(a); 4.22, 1H, m, Tyr□; 4.02, 1H, br, NCH₂N(b); 3.56, 1H, bd, J=17.2 Hz, Gly□(a); 3.48, 1H, m, TyrC'H; 3.29, 1H, bd. J=17.2 Hz, Gly \square (b); 2.57-2.88, 4H, e, Tyr \square CH₂ and \square -aldehyde CH₂; 2.22, s, H₂O; \square 1.48-1.08 (1.20 peak), 9H, vbr, Boc 3xCH₃. ¹³C NMR (75 MHz, CD₃CN): □ 201.9; 171.4; 158.7; 154.3; 139.0; 137.6; 132.6; 131.9, 129.92, 129.85, 129.6, 129.2, 128.9, 116.0: ArCH; 80.41 (Boc tert.); 70.99 (CH₂); 67.62 (br, CH₂); 67.44 (br, CH₂); 60.29 (2xCH, co-incident peaks determined by comparative intensity); 52.99 (br, CH₂); 43.58 (br, CH₂); 35.94 (br, CH_2); 28.78 (br, $Boc 3xCH_3$).

Example (C). Synthesis of \Box -turn mimetics I(i) for the Gly-Phe-Leu sequence by the short method (which can be used when R^1 = hydrogen)

20

Preparation of 69:

5

10

15

20

Boc-glycine was coupled with N,O-dimethyl hydroxylamine hydrochloride, 1 equivalent, in DMF/CH₂Cl₂ (1:5) using HBTU reagent (1 eq.) and DIEA (2 eq.) at room temperature. The CH₂Cl₂ was evaporated in vacuo and the residue partitioned between diethyl ether and aq. NaHCO₃. The aqueous layer was separated and the ether layer washed in turn with 1M HCI (x2), aq. NaHCO₃, brine, and then dried over MgSO₄. Filtration and removal of the solvent in vacuo left the product amide 69 as a viscous oil that slowly crystallised to a waxy solid and was further purified by chromatography on silica gel. Yield was >90%. ¹H NMR (300 MHz, CDCl₃): □ 5.3, 1H, bs, NH; 4.09, 2H, bd, □H₂; 3.72, 3H, s, OCH₃; 3.20, 3H, s, NCH₃; 1.46, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃): □ 79.6; 61.4; 41.7; 32.4; 28.3.

Preparation of 70:

A solution of 11.6 g (53 mmol) of Boc-glycine N,O-dimethylhydroxylamide in dry THF (70 mL) under nitrogen in a 250 mL round bottom flask was stirred and cooled in an ice bath. To this was added vinyl magnesium bromide in THF (~120 mmol of a 1M solution) by

10

15

20

25

30

syringe over 10 minutes. The solution was stirred for 2 h and then quenched by pouring into a mixture of crushed ice and 1M HCI which was then extracted with CH_2Cl_2 (x2). The organic extracts were washed with water/brine (x2), aq. NaHCO₃ and water/brine followed by drying over MgSO₄. Evaporation of the solvent left 9.6 g of a mobile oil (98% crude) which by NMR was ~95% the ketone product **70**. This material was used without further purification in the conjugate addition step. ¹H NMR (300 MHz, CDCl₃): \Box 6.37, 2H, m (ABX, Jab=2.5 Hz, Jax/bx=9.0, 17.5 Hz), vinyl CH₂; 5.95, 1H, dd, J=2.5, 9.0 Hz, vinyl CH; 5.37, 1H, bs, NH; 4.26, 2H, d, J=4.6 Hz, glycyl \Box H₂; 1.46, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃): \Box 194.9 ketone; 155.8 carbamate; 133.6 vinyl; 129.6 vinyl; 79.8 tBoc; 48.32 Gly \Box ; 28.28 Boc. Preparation of **71**:

To a solution of 3.0 g (~15 mmol) of crude 70 in THF (40 mL) was added 3.4 g of leucine methyl ester hydrochloride (~1.2 eq) and 2.4 g (1.2 eq) of diisopropylethylamine. After 2 h the reaction was diluted with ether (200 mL) and extracted with cold 1M HCI (3x50 mL) (discard this ether layer). The aq. extracts were immediately neutralised with solid NaHCO3 and this solution was then back extracted with ether, and the ether washed with water (x3) and finally brine and dried over MgSO₄. Evaporation of the solvent left ~5.3 g of product 71 as an oil with very good purity, contaminated with a small amount of leucine methyl ester. Flash chromatography to separate the product was not very successful as the amine and amino ketone tended to co-elute. TLC EA/LP Rf=0.35. 1H NMR (300 MHz, CDCl₃):

5.36, 1H, bm, NHBoc; 4.03, 2H, d, J=5 Hz, Gly□; 3.72, 3H, s, OCH₃; 3.26, 1H, t, J=7.5 Hz, Leu□; 2.93, 1H, dt, J=12, 6 Hz; 2.72, 1H, dt, J=12, 6 Hz; 2.50, 2H, m; 2.0, 1H, bs, NH; 1.69, 1H, m, Leu□; 1.45, 11H, m, Boc(9H) and Leu□(2H); 0.90, 6H, m, Leu□. ¹³C NMR (75 MHz, CDCl₃): □ 205.1; 176.1; 155.5; 79.8 tBoc; 60.04; 51.64; 50.53; 42.63; 42.57; 40.55; 28.26 Boc; 24.81; 22.63; Mass Spectrum (ISMS) m/z 331.4 (M+H+), calculated for 22.17. C₁₆H₃₀N₂O₅: 330; fragments (OR 60): 275.2 (-tBu).

10

15

20

25

30

Preparation of 72:

The amine 71 was protected as the benzyl carbamate by standard procedures as follows: the crude amine product 71 (1.68 g, ~5 mmol) was dissolved in ethyl acetate (30 mL) to which was added a solution of KHCO₃ (1.2 g) in water (15 mL). This mixture was vigorously stirred and cooled in an ice bath and to it was added benzyl chloroformate (780 uL of a 95% solution, 5.2 mmol) dropwise over 5 min. The reaction was stirred for a further 15 min then allowed to warm to room temperature with stirring for an additional 2 h. After this time the mixture was diluted with ether (100 mL), the aqueous layer seperated, and the organic layer washed with 1M HCl, aq. NaHCO₃, brine and then dried over MgSO₄. Evaporation of the solvent left ~2.6 g crude oil which was purified by flash chromatography eluting with 25%EtOAc in light pet; combination of the main fractions gave a yield of 86% (2.02 g) of 72. TLC EA:2LP Rf=0.56. NMR signals split due to amide rotamers (~1:1) are placed in parentheses where possible. ¹H NMR (300 MHz, CDCl₃): □ 7.40-7.23, 5H, Ar; 5.28-5.02, 3H, m's, $CH_2Ph + NH$; (4.64, m, 4.43, m) 1H; (3.98, bs, 3.88, bs) 2H; 3.72-3.51, 4H, includes (3.67, s, 3.55, s) OCH₃ + 1H; 3.45, 1H, m; 2.78, 2H, m; 1.75, 2H, m; 1.53, 1H, m; 1.43, 9H, s, Boc; 0.91, 6H, m, Leu□ CH₃x2. ¹³C NMR (75 MHz, CDCl₃): □ (204.9, 204.5) ketone; (172.5, 172.3) ester; (156.1, 155.8) carbamate; 155.6, carbamate; (136.2, 136.0) ipso; 128.5, 128.2, 128.1, 128.0: ArCH; 79.80, tBoc; 67.48; (58.50, 58.32); 52.12; 50.30; (41.37, 39.87, 39.78, 38.87, 38.60, 37.98) 3C; 28.23, Boc; (24.83, 24.67); 23.09; (21.46, 21.39). Mass Spectrum (ISMS) m/z 465.3 (MH+), calculated for $C_{24}H_{36}N_2O_7$: 464; fragments (OR 70): 409.2, (-tBu); 365.2, (-Boc).

Preparation of amines 73:

To a solution of **72** (700 mg, 1.5 mmol) in 15 mL of 1,2-dichloroethane was added phenylalanine benzyl ester p-toluene sulfonate (900 mg, 2.1 mmol) and sodium triacetoxy borohydride (850 mg, 4.0 mmol). The mixture was stirred at room temperature for 24 h and then the solvent removed under vacuum and the residue partitioned between ethylalanine partitioned partitioned between ethylalanine partitioned partitioned partitioned between ethylalanine partitioned partitio

10

15

20

25

acetate and aq. NaHCO3, the aqueous layer separated, and the organic layer washed with water then brine and then dried over MgSO4. Evaporation of the solvent left 1.2 g crude oil which was purified by flash chromatography eluting with 25-40% EtOAc in light petroleum ether to give a yield of 76% (800 mg) of the product (a clear oil). The product diastereomers 73 were not seperable under these chromatography conditions. TLC 40%EA in LP Rf=0.48. ¹H NMR (300 MHz, CD₃CN): □ (not very informative due to the presence of rotamers/diastereomers) 7.45-7.05 aromatic protons; (5.46 m, 5.31 m)~1/2H; 5.15-5.00, ~4H, m, OCH₂Ph; 4.95, ~1/4H, m; (4.51, m, 4.37, m): 1H; 3.85-3.10, ~5H, e (including 3.63, s, 3.58, s: 3H, OCH₃); 3.10-2.70, 5H, e; 2.45 broad water peak; 1.80-1.45, 5H, m's; 1.40, 9H, s, Boc; 0.90, 6H, bs, Leu ... 13C NMR (75 MHz, CD₃CN):

(signals are grouped in parentheses where they can be reasonably assigned to equivalent carbons in the different diastereomers/rotamers) (175.6, 175.4(br)); 173.6; 157.2 (br); (139.0, 139.2, 138.5, 138.3, 137.3) 3x ipso; 130.8, 130.7 129.9 129.71, 129.66, 129.3, 129.0, 128.0: Ar CH; (79.87, 79.62) Boc tertiary; 68.22 (CH₂, OBn); 67.75 (CH₂, OBn); (61.67, 61.55) (CH); 59:39 (CH); (56.51, 55.82, 55.61) (CH); 53.11 (OCH₃); (45.56, 45.16, 44.73, 44.61, 44.43, 44.24, 43.42, 43.04) (2xCH₂); (40.77, 40.15, 40.03, 39.42, 39.27) (2xCH₂); (39.66, 32.60, 32.45, 31.44) (CH₂); 29.04 (CH₃ Boc); 29.93 (CH); 23.88 (CH₂); 22.36 (CH₂). Mass Spectrum (ISMS) m/z 704.4 (M+H+), calculated for C₄₀H₅₃N₃O₈: 703.

Preparation of 74 and 75:

74 (R) - major product 75 (S) - minor product

The mixture of epimeric amines **73** (260 mg, 0.4 mmol) was dissolved in methanol (20 mL) and 10% palladium on carbon added (100

CO₂CH₃

10

15

20

25

30

mg). The solution was hydrogenated (40 psi H₂) at room temperature for 3 h to give the deprotected amino acid (MH+=480Da). After filtration, the solvent was removed and the residue (170 mg) was dissolved in DMF (5 mL) and diluted with CH₂Cl₂ (50 mL). To this solution was added HBTU (180 mg, 0.48 mmol) and DIEA (150 mg, 1.2 mmol). After stirring for 10 min at room temperature the solution was diluted with aq.NaHCO3, the aqueous layer separated, and the organic layer washed with water (x3) then brine and then dried over MgSO₄. Evaporation of the solvent left an oil which was purified by flash chromatography eluting with 20-40% EtOAc in light petroleum ether. The product diastereomers were just separable under these conditions, with the minor diastereomer 75 eluting first to give a yield of 18% (30 mg) followed by the major diastereomer 74 in 50% (85 mg) yield. TLC EA:LP 1:1 Rf=0.43, 0.29. 1H NMR (300 MHz, CD₃CN): □□ Isomer 75: 7.29, 4H, m, ArH; 7.22, 1H, m, ArH; 5.17, 1H; dd, J=6.5, 8.4 Hz; 5.08, 1H, m; 3.65, 3H, s, OCH₃; 3.61, 1H, dd, J=11.4, 15.6 Hz; 3.27, 1H, ddd, J=1.5, 5.7, 15.9 Hz; 3.12, 1H, dd, J=4.5, 14.3 Hz; 2.98, 1H, bm; 2.72, 1H, m; 2.64, 1H, dd, J=9.9, 14.3 Hz; 2.57, 1H, bm; (2.17, H_2O); 1.68, 3H, m; 1.60, 1H, m, Leu \Box ; 1.36, 9H, s, Boc; 1.16, 1H, m; 0.95, 3H, d, J=6.4 Hz, Leu□; 0.93, 3H, d, J=6.6 Hz. isomer 74: 7.29, 4H, m, ArH; 7.22, 1H, m, ArH; 5.11, 1H, dd, J=5.6, 9.4 Hz; 4.29, 1H, br, NHBoc; 3.81, 1H, dd, J=4.6, 9.8 Hz; 3.65, 3H, s, OCH₃; 3.59, 1H, dd, J=10.8, 15.2 Hz; 3.19, 1H, dd, J=5.5, 15.2 Hz; 3.13, 1H, dd, J=4.5, 13.8 Hz; 2.94, 2H, m's; 2.71, 1H, m; 2.64, 1H, dd, J=10.3, 13.3 Hz; (2.17, H₂O); 1.76, 1H, m; 1.69, 2H, m; 1.57, 2H, m; 1.36, 9H, s, Boc; 0.93, 6H, d, J=6.5 Hz. ¹³C NMR (75 MHz, CDCl₃):

Isomer 75 (5S): 175.2; 172.5; 155.9; 138.9; 129.3; 128.5; 126.4; 79.2; 60.91; 60.62; 55.65; 52.19; 45.70; 43.98; 38.12; 37.99; 33.46; 28.30, Boc; 25.01; 23.10; 21.93. Isomer 74 (5R): 175.1; 172.5; 155.7; 139.3; 129.3; 128.7; 126.8; 78.9; 56.01; 55.80; 53.05; 52.14; 42.07; 40.70; 38.01; 37.98; 31.51; 28.26, Boc; 25.03; 23.11 21.74. Mass Spectrum (ISMS) m/z 462.3 (MH⁺), calculated for C₃₂H₄₅N₃O₅: 461 fragments (OR 70): 406.2 (-tBu).

20

25

Example (D). Selective synthesis of the 3(S), 5(S) diastereomer 75 by the short method

The 3(S)5(S) diastereomer, the minor product formed as described above, can be selectively synthesised by the use of an intramolecular reductive amination-cyclisation as described below:

10 Preparation of acyl fluoride 76:

Z-phenylalanine acid fluoride was prepared by general literature methods (Carpino *et al.*, 1990; Wenschuh *et al.*, 1994) as follows: 1.1 equivalents of diethylaminosulfurtrifluoride (DAST) were added to ZPheOH in dry dichloromethane solution under nitrogen at 0°C. After stirring for 15 min the reaction was worked up by pouring onto iced water and separating the organic layer, washing once with cold water and then drying over MgSO₄. The product was purified by precipitation from ether/petroleum ether and dried *in vacuo*. ¹H NMR (300 MHz, CDCl₃): □ 7.36, 8H, m's; 7.28, 2H, m; 5.30, 1H, bd; J=7.5 Hz, NH; 5.13, 2H, s, OCH₂Ph; 4.85, 1H, m, □H; 3.20, 2H, m, □H₂. ¹³C NMR (75 MHz, CDCl₃): □ 161.8, d, ¹J_{CF}=370 Hz; 155.5; 135.7; 134.2; 129.1; 129.0; 128.5; 128.3; 128.1; 127.7; 67.36; 53.50, d, ²J_{CF}=59 Hz; 36.70. Preparation of 77:

To the amine **71** (2.7 g, 8.2 mmol) dissolved in CH₂Cl₂ (40 mL) was added Z-phenylalanine acid fluoride **76** (prepared as described above) (3.0 g, 10 mmol) and DIEA (1.3 g, 10 mmol) and the solution stirred at room temperature under nitrogen for 30 h. The solvent was then evaporated *in vacuo* and the residue dissolved in ether and

الدين و يشور.

5

10

15

25

extracted in turn with 1M HCI (x2), 10% aq. Na₂CO₃ (x2), then brine and then dried over MgSO₄. The solution was filtered and the solvent removed in vacuo. The resulting oil was purified by flash chromatography eluting with 20-40% ethyl acetate in light petroleum ether for a yield of about 80% of the target 77 as a clear oil. TLC 40%EA:LP Rf=0.40. 1H NMR (300 MHz, CDCl₃):

7.41-7.13, 10H, Ar; 5.48, 1H, bd, J=9.2 Hz, NHCbz; 5.19, 1H, bm, NHBoc; 5.09, 2H, s, OCH₂Ph; 4.76, 1H, dt, J=6.4, 8.9 Hz, Phe□; 4.38, 1H, dd, J=5.2, 9.3 Hz, Leu□; 3.92, 2H, d, J=4.5 Hz, Gly□; 3.60, 3H, s, OCH₃; 3.54, 1H, m; 3.38, 1H, m; 3.08, 1H, dd, J=8.4, 13.3 Hz; 2.93, 1H, dd, J=6.1, 13.1 Hz; 2.65, 2H, m; 2.80, 1H, m; 2.64, 1H, m; 1.46, 9H, s, Boc; ~1.38, 1H, m; 0.90, 6H, 2xd, J=6.6, 6.5, Leu[]. 13C NMR (75 MHz, CDCl₃) amide rotamers (~5:1): only the major peak of rotamer peak pairs is reported:

204.1; 172.1; 171.4; 156.7; 155.6; 136.2; 135.8; 129.4-127.1; ArCH; 79.8; 66.82; 58.15; 52.25; 52.05; 50.28; 41.32; 39.58 (2 coincident signals as determined by relative intensity, shift and the presence of both minor rotamer peaks); 37.82; 28.23, Boc; 24.67; 23.08; 21.67. Mass Spectrum (ISMS) m/z 612.3 (M+H+), calculated for C₃₃H₄₅N₃O₈: 611; fragments: (OR 60): 556.3 (-tBu); 512.3 (-Boc).

20 Selective preparation of **75** from **77**:

The ketone 77 (1mmol) was dissolved in 0.1M methanolic HCI (30ml) and 10% palladium on activated carbon (200mg) was added. The solution was hydrogenated at 30 psi H₂ (room temperature) for 8 h and then diluted with aq. NaHCO₃ and extracted with ethyl acetate. The organic layer was washed with water (x2) and then brine then dried over MgSO₄. Filtration and removal of solvent in vacuo left the crude product 75 in good yield and purity. Analysis of the crude product by NMR and by TLC did not reveal any of diastereomer 74. The reaction was estimated to be >95% stereoselective.

30 Example (E). Synthesis of a biologically active □-turn mimetic for the Arg-Gly-Asp sequence

.4

. -

÷ 5-

Preparation of 78:

5

10

15

The \Box , \Box -unsaturated ketone **70** (1.0 g, 5.4 mmol, prepared as previously described) was reacted with phenethylamine hydrochloride (1.07 g, 6.8 mmol) and DIEA in THF by the method previously described for the preparation of **71**. The crude product **78** was used without further purification for the next reaction. Mass Spectrum (ISMS) m/z 307.2 (MH+), calculated for $C_{17}H_{26}N_2O_3$: 306; fragments (OR 60): 250.9 (-tBu). Preparation of **79**:

To a stirred solution of Boc-aspartic acid □-benzyl ester (3.23 g, 10 mmol) in CH₂Cl₂ (10 mL) was added dicyclohexylcarbodiimide (10 mL of 0.5M solution in CH₂Cl₂) at room temperature. A copious precipitate of dicyclohexylurea soom formed; after 10 min the solution was filtered, and the solvent removed *in vacuo*. The residual oil was

10

15

added to a solution of crude 78 (1.3 g) in THF, followed by DIEA (645 mg, 5 mmol), and the solution stirred for 4 h. The reaction mixture was diluted with ether/ethyl acetate and washed with 1M HCl, aq. NaHCO3, water, brine and dried over MgSO₄. The crude product was purified by flash chromatography eluting with 30-50% ethyl ether in petroleum ether to give a reasonable yield of 79 (estimated as 80% based on 78) as a clear oil. ¹H NMR (300 MHz, CDCl₃, amide rotamers present): □□ 7.38-7.16, 10H, m, Ar; 5.37, 1H, bd, J=9 Hz, AspNHBoc (minor rotamer 5.33, J=10 Hz); 5.25, m, 1H (Gly NH); 5.10, 2H, m, OCH₂Ph; 4.89, 1H, m; 3.93, 2H, d, J=4.4 Hz, Gly0; 3.67-3.53, 3H, m's; 3.47, 1H, m; 2.95-2.52, 6H, m's (including 2.88, 2H, m; 2.63, 2H, ABX, J=15.8, 7.3, 5.8 Hz, \Box H₂Asp); 1.44, 18H, multiple singlets, 2xBoc. ¹³C NMR (75 MHz, CDCl₃): [(major rotamer only) 204.7; 171.0; 170.3; 155.6; 154.8; 137.7; 135.5; 128.9, 128.6, 128.5, 128.2, 126.6; ArCH; 80.06; 79.73 (2x tBoc); 66.57; 50.55; 50.33; 46.99; 42.24; 37.69 (2 signals); 35.50; 28.22 (2x Boc). Mass Spectrum (ISMS) m/z 612.3 (MH+), calculated for $C_{33}H_{45}N_3O_8$: 611 fragments (OR 60): 556.1 (-tBu); 512.1 (-Boc). Preparation of 80:

The ketone 79 (390 mg, 0.64 mmol) in CH_2Cl_2 (2 mL) was treated with trifluoroacetic acid (2 mL) and the solution stirred for 30 min 20 at room temperature. The volatiles were then removed in vacuo and CH₂Cl₂ (3 mL) added and removed in vacuo (x2). The residual oil was dissolved in 1,2-dichloroethane (5 mL) and NaBH(OAc)₃ (270 mg, 1.3 mmol) added. The mixture was stirred for 20 min then the solvent removed and the residue dissolved in ethyl acetate and washed with aq. 25 Na₂CO₃ and then brine and then dried over MgSO₄. The crude product 80 (after solvent removal 210 mg, 84%) was of good purity by MS and NMR, with only one diastereomer observed (>95% diastereoselectivity). 1H NMR (300 MHz, CDCl₃):

7.39-7.10, 10H, m, Ar; {5.20, 5.16, 5.14, 5.10}, 2H, ABq, J=12.5 Hz) OCH₂Ph; 3.86, 1H, t, J=6.3; 3.76-3.43, 3H, 30 m's; 3.14, 1H, bdd, J=15, 5 Hz; 2.98-2.76, 5H, e; 2.70, 1H, dd, J=7.4, 16 Hz; 2.46, 1H, m; 1.64, 1H, bm; 1.06, 1H bm. 13C NMR (75 MHz,

ij

...

. .

5

10

15

20

30

CDCl₃): ☐ 173.9; 172.0; 138.9; 135.9; 128.7, 128.4, 128.0, 126.3: Ar; 66.16; 60.49; 56.55; 51.24; 48.39; 45.14; 38.05; 34.15; 33.01. Mass Spectrum (ISMS) m/z 396.2 (MH⁺), calculated for C₂₃H₂₉N₃O₃: 395. Preparation of **81**:

The crude amine product 80 (140 mg, ~0.35 mmol) was coupled with BocArg(Tos)OH (182 mg, 1.2 eq) using the BOP reagent (188 mg) and DIEA (55 mg) in DMF/CH₂Cl₂ (5ml). The CH₂Cl₂ was evaporated in vacuo and the residue partitioned between diethyl ether/ethyl acetate and aq. NaHCO3. The aqueous layer was separated and the organic layer washed in turn with 1M HCl (x2), water (x2), aq. NaHCO3, brine, and then dried over MgSO4. Filtration and removal of the solvent in vacuo left the crude product amide 81 which was purified by flash chromatography eluting with 5-10% ethanol in ethyl acetate (yield 260 mg, 90%). TLC 10% EtOH in EtOAc Rf=0.38. 1H NMR (300 MHz, CD₃OD):

7.74, 2H, d, J=7 Hz; 7.4-7.15, 12H, m's; 5.15, 2H abq, J=11 Hz, OBn; 4.26, 1H, m; 4.03, 1H, m; 3.73, 2H, m; 3.48-3.07, 7H, e; 3.07, 1H, m; 2.92-2.73, 3H, m's; 1.92, 1H, m; 1.73, 1H, m; 1.66-1.45, 4H, e; 1.42, 9H, s, Boc. ¹³C NMR (75 MHz, CD₃OD): □ 176.1; 172.5; 172.0 (br); 158.8; 158.1; 143.7; 142.2; 140.3; 137.5; 130.4; 130.1; 129.72; 129.68; 129.4; 128.4; 127.6; 127.3; 127.2; 80.92 (t); 67.75 (CH₂); 62.55 (CH); 57.27 (CH); 56.00 (CH); 52.55 (CH₂); 48.74 (CH₂); 44.42 (CH₂); 41.22 (br, CH₂); 37.00 (CH₂); 35.10 (CH₂); 32.41 (CH₂); 30.15 (CH₂); 28.87 (Boc CH₃); 27.24 (br, CH₂); 21.57 (CH₃). Mass Spectrum (ISMS) m/z 806.4 (MH $^+$), calculated for C₄₁H₅₅N₇O₈S: 805.

25 Preparation of 82:

The amine 81 (50 mg, 0.06 mmol) in THF (0.6 mL) was cooled in a dry ice acetone bath and ammonia gas added until ~30 mL of ammonia had condensed. Small pieces of sodium metal (3-6 mg) were added until the blue colour persisted. The reaction was quenched by the addition of ammonium carbonate (25 mg), the dry ice bath removed and the solvent allowed to evaporate at room temperature. The residue (which gave a crude mass spectrum with the product mass as the only)

10

15

20

25

significant peak) was purified by reversed phase HPLC (Vydac C18) eluting with 85% solvent A (=0.1% CF_3COOH in H_2O):15% solvent B (=0.1% CF_3COOH and ~10% H_2O in CH_3CN) for 2 minutes followed by a 2%/min gradient. Only one product diastereomer was observed in the HPLC traces. Mass Spectrum (ISMS) m/z 562.3 (M+H⁺), calculated for $C_{27}H_{43}N_7O_6$.

Preparation of 83:

The amine 81 was dissolved in CH₂Cl₂/CF₃CO₂H (2ml, 1:1) and stirred at room temperature for 30 minutes after which the Boc group had been removed. 10ml of CH₂Cl₂ was then added and the volatiles removed in vacuo (repeat once). The residue was again dissolved in (2 eq.) added along with acetic anhydride and CH₂Cl₂ diisopropylethylamine (DIEA, 5 eq.), and the reaction stirred at room temperature for 2 h. The volatiles were removed in vacuo and the residue dissolved in ethyl acetate and washed with aq. NaHCO3 then brine and then dried over MgSO₄. Filtration and removal of the solvent in vacuo left_ the crude product 83 as an oil in reasonable purity. The ¹H NMR was badly broadened in common solvents at room temperature. 13C NMR (75MHz, CDCl₃):

173.7; 172.4; 171.9; 171.0; 157.0; 142.1; 140.4; 138.8; 135.8; 129.2, 128.7, 128.4, 128.1, 128.0, 126.3, 125.8: ArCH; 66.22, OCH₂Ph; 60.08, CH; 56.09, CH; 52.94, br, CH; 51.06, CH₂; 48.21, CH₂; 44.31, CH₂; 40.13, br, CH₂; 37.79, CH₂; 34.16, CH₂; 32.97, CH₂; (29.59, 29.50) 1C, br, CH₂; 25.64, br, CH₂; 22.91, CH₃; 21.32, CH₃. Mass Spectrum (ISMS) m/z 748.2 (MH+), calculated for C₃₇H₄₉N₇O₇S: 747.

Preparation of 84:

Compound 84 was prepared from 83 by dissolving metal reduction as described for the preparation of 82 above. Purification was carried out by HPLC under the same conditions as for 82.

30 <u>Testing of Arg-Gly-Asp mimetics</u> 82 and 84 for inhibition of platelet aggregation in human platelet rich plasma (PRP)

10

15

20

The peptide sequence arginine-glycine-aspartic acid (RGD) is important to the binding of proteins to certain integrin receptors, such as the GP_{IIb-IIIa} receptor found on the surface of platelets. Several cyclic peptides having the RGD sequence have been found to antagonise the binding of plasma proteins to the GP_{IIb-IIIa} receptor, thereby inhibiting blood clotting. GP_{IIb-IIIa} antagonists have therapeutic potential as anti-thrombotics, there are several in early clinical trials(Humphries, Doyle *et al.*, 1994). Mimetics based on □-turn structures centred on the Asp residue have been successful, this structure was chosen to test the compounds of the invention.

Solutions of the compounds to be tested were made up in water. Platelet aggregation induced by adenosinediphosphate (ADP, 10 M) in human PRP was measured by the decrease in light scattering on aggregation measured with a platelet aggregometer. The tetrapeptide Ac-Arg-Gly-Asp-Ser-NH2 was used as a positive control.(Callahan et al., 1992) Compounds 82 and 84 were both found to inhibit platelet aggregation in a dose dependent manner, and both exhibited stronger inhibition than the control peptide. Compound 84 was the strongest, having inhibitory activity approximately five times more potent than Ac-Arg-Gly-Asp-Ser-NH2 under the conditions of the test.

Example (F). Synthesis of fully substituted \(\pi\)-turn mimetics for the Phe-Leu-Ala sequence in both the 4(R) and 4(S) configurations

The synthesis up to the final common intermediate for the 4(R) and 4(S) diastereomers, the aldehyde 93, is summarised below:-

10

15

N,O-dimethylhydroxylamide 85 Bocphenylalanine synthesised by the general solution phase coupling procedure as N,O-dimethyl Boc-phenylalanine and described from previously Purification: on a Yield: ~quantitative. hydroxylamine hydrochloride. short silica column eluting with ether. ¹H NMR (300 MHz, CDCl₃): [] 7.33-7.12, 5H, m, Ar; 5.20, 1H, bd, J~7 Hz, NH; 4.95, 1H, bm, Phe□; 3.66, 3H, s, OCH₃; 3.17, 3H, s, NCH₃; 3.06, 1H, dd, J=6, 13.5 Hz, Phe□; 2.88, 1H, dd, J=7.5, 13.5 Hz; 1.40, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃):
□ 172.2; 155.1; 136.5; 129.4; 128.2; 126.7; 79.5; 61.4; 51.4; 38.8, Phe D; 32.0; 28.2, Boc.

The amide 85 was reduced to Bocphenylalanine aldehyde 86 by the method of Fehrentz and Castro (Fehrentz and Castro, 1983)

Briefly: amide (2 mmol) dissolved in dry ether (20 mL) and cooled and in

10

15

20

25

30

an ice bath under nitrogen, then LiAlH₄ (95 mg, 2.5 mmol) added and stirring continued 15 min. Then KHSO₄ (477 mg, 3.5 mmol) in 10 mL water added and then 150 mL ether and wash with 1M HCl (cold) (x3), aq. NaHCO₃, brine, and dried over MgSO₄. Removal of the solvent left the solid aldehyde in ~90% crude yield containing some of the overreduced alcohol as the only significant impurity. TLC EtOAc:light pet. Rf=0.5. ¹H NMR (300 MHz, CDCl₃): □ 9.63, 1H, s, aldehyde; 7.37-7.13, 5H, m, Ar; 5.07, 1H, bs, NH; 4.43, 1H, m, Phe□; 3.11, 2H, d(AB) Phe□; 1.43, 9H, s, Boc. ¹³C NMR (75 MHz, CDCl₃): □ 199.4, aldehyde; 155.3, carbamate; 135.7, ipso; 129.3, 128.7, 127.1: ArCH; 80.2, tBoc; 60.8, Phe□; 35.5, Phe□; 28.2, Boc

Methyl leucinate hydrochloride (0.80 g, 4.4 mmol) was neutralised with 10% aq. Na₂CO₃ solution (25 mL), and the solution was mixed with brine (25 mL) and extracted with CH₂Cl₂ (3x20 mL). The organic extracts were dried over MgSO₄ and most of the solvent removed under vacuum (~2 mL residue). This solution of methyl leucinate was added to Boc phenylalanine aldehyde 86 (1.1 g, 4.4 mmol) in CH₂Cl₂ (5 mL), the stirred solution soon became turbid due to the separation of water, dried MgSO₄ (500 mg) was added and the solution cleared. After 30 min the solution was filtered into a dried flask under nitrogen. NMR analysis showed that all the aldehyde had been converted to the imine 87 and that significant racemisation had not taken place. The imine was used without further purification for the allylation reaction. ¹H NMR (300 MHz, CDCl₃):

7.61, 1H, d, J=1.3 Hz, imine; 7.32-7.14, 5H, m, Ar; 5.69, 1H, bd, J=4.5 Hz, NH; 4.49, 1H, m, Phe ; 3.85, 1H, dd, J=5.5, 8.5 Hz, Leu□; 3.69, 3H, s, OCH₃; 3.20, 1H, dd, J=5.0, 14.5 Hz, Phe□; 2.96, 1H, dd, J=8.0, 13.5 Hz; Phe: 1.63, 1H, m; 1.46, 9H, s, Boc; 1.42, 1H, m; 1.30, 1H, m; 0.88, 3H, d, J=6.5 Hz, Leu\(\text{L}\); 0.80, 3H, d, J=6.5 Hz, Leu□. ¹³C NMR (75 MHz, CDCl₃): □ 171.7, ester; 164.3, CH, imine; 154.6, carbamate; 136.1, ipso; 128.9, 127.7, 126.0: ArCH; 78.56, tBoc; 69.51; 54.08; 51.32; 41.02, CH₂; 38.04, CH₂; 27.73, Boc; 22.35; 22.48; 20.63.

10

15

20

25

B-allyl-9-borabicyclononane Rg1a can be synthesised from (synthesised in turn from B-methoxy-9-borabicyclononane methanolysis of 9-BBN (Kramer and Brown, 1974)) by the method of Kramer (Kramer and Brown, 1977). Alternatively the following one-pot synthesis from 9-BBN was used: a suspension of 9-BBN (crystalline dimer, 8.97 g, 73.5 mmol) in anhydrous ether (75 mL) was stirred under nitrogen and cooled to 0°C. Methanol (3.3 mL, 81 mmol) was slowly added by syringe (gas evolved), and vigorous stirring continued for ~3 h (9-BBN gradually dissolves, gas evolution ceases). Allylmagnesium bromide in ether (81 mL of a 1.0M solution) was slowly added to the solution (still cooled to 0°C); (a thick grey ppt. forms, stirring may be difficult). Stirring was continued for 1 h then the solution was allowed to warm to room temperature and the ether was pumped off under moderate vacuum (~300->20mbar). The residue was re-suspended in anhydrous hexane (100 mL) and then stirring stopped to allow the magnesium salts to settle out. The solution was estimated by reaction with a known amount of methylphenylketone in ether (found to be ~0.57M, equal to 78% yield). The clear solution of B-allyl-9-BBN was used directly for allylation of the imines. (This procedure was adapted from one described by Rachlera and Brown (Racherla et al., 1992)) The imine 87 (~23 mmol) was dissolved in dry diethylether (100 mL) under nitrogen and the stirred solution cooled to -78°C. B-allyl-9-BBN (47.5 mL of ~0.57M solution in hexane, ~27 mmol) was added and the solution stirred for 1 h and then allowed to warm to room temperature with stirring for an additional 1 h. Glacial acetic acid (1.5 mL) was added and the ether was removed in vacuo. The residue was dissolved in acetonitrile (100 mL) and more glacial acetic acid (5 mL) added. The solution was then refluxed until all of the borane adduct had been converted to the amine (~24 h, monitored by TLC: Rf adduct>Rf amine = 0.32 in 1:5 EtOAc:light pet.). acetonitrile was removed in vacuo and the residue partitioned between 30 ether/light petroleum and 10% aq. Na₂CO₃. The organic layer was washed again with 10% aq. Na₂CO₃ and then extracted with a solution of

....

10

15

20

25% methanol in 0.5M HCI (three times), the organic layer containing the neutral reaction products (~6 g) was discarded. The ag. acid extracts were immediately neutralised with solid NaHCO3 and then extracted with ether. The ether solution was washed with water then brine and then dried over MgSO₄. Evaporation of the solvent left the amine products (5.9 g) which were further purified by flash chromatography eluting with 7.5-15% ethyl acetate in light petroleum for a yield of 50+% of the amines 88 based on the crude aldehyde 86 used in the imine formation. Some separation of the diastereomers was observed in the chromatography, but they were not well resolved. Alternatively the crude amines were hydrolysed to the amino acid as described below and purified by recrystallisation. ¹H NMR (300 MHz, CDCl₃), major diastereomer: © 7.32-7.13, 5H, m, Ar; 5.84, 1H, m, vinylCH; 5.11, 2H, m, vinylCH₂; 5.00, 1H. d. J=8 Hz, NHBoc; 3.88, 1H, m, Phe ; 3.66, 3H, s, OCH₃; 3.40, 1H, t, J=7 Hz, Leu0; 2.87, 1H, dd, J=5, 13 Hz, Phe0; 2.69, 2H, m's: Phe + CH(homoallyl); 2.23, 2H, m, allyl; 1.7, 1H, b, NH(amine); (1.65, 1H, m; 1.47, 2H, m) Leu□+□; 1.33, 9H, s, Boc; 0.90, 6H, t(2 doublets) J=7, 7 Hz, Leu□. ¹³C NMR (75 MHz, CDCl₃), major isomer: □ 176.1; 155.4; 138.6, ipso; 135.2, CH vinyl; 129.2, 128.2, 126.1; CHAr; 117.4, CH₂ vinyl; 78.8, tBoc; 58.94; 58.56; 54.10; 51.71; 42.87; 36.52; 35.61; 28.24, Boc; 24.78; 22.68; 22.23. Mass Spectrum (ISMS) m/z 419.2 (MH+), calculated for C₃₂H₄₅N₃O₅: 418 fragments (OR 65): 363.2, (-tBu).

in methanol/water and LiOH.H₂O (800 mg, 19 mmol) added. The solution was stirred at room temperature until the hydrolysis was complete (12 h) and then neutralised with 1M HCl (19 mL). On standing a copious white precipitate formed which was filtered off and washed with water. The solid was recrystallised from ethanol-water (~95:5) to give fine needles of (mainly) the major diastereomer 89 (first crop 1 g), m.p.:175-177°C. The product was further recrystallised as required. 1H NMR (300 MHz, CD₃OD): (ref. 3.31 ppm) 7.33-7.18, 5H, m; 5.90, 1H, m; 5.35, 1H, discontinuous contents.

10

15

20

25

30

J=17.1 Hz; 5.26, 1H, d, J=10.2 Hz; 4.31, 1H, m; 3.65, 1H, dd, J=5.7, 7.9 Hz; 3.27, 1H, m; 2.92, 1H, dd, J=5.2, 14.0 Hz; 2.76, 1H, dd, J=10.1, 14.0 Hz; 2.59, 1H, m; 1.82, 1H, m; 1.37, 9H, s, (Boc); 0.97, 3H, d, J=7 Hz; 0.94, 3H, d, J=7 Hz. 13 C NMR (75 MHz, CD₃OD): \Box (ref. 49.15 ppm) 173.7; 159.4; 138.8; 134.5; 130.33; 129.8; 128.0; 120.5; 81.34; 63.65; 55.84; 41.19; 37.90; 32.70; 28.78; 26.11; 23.56. Mass Spectrum (ISMS) m/z 405 (MH+), calculated for $C_{23}H_{36}N_2O_4$: 404.

The amino acid 89 was esterified to 90 by the method of Bodansky and Bodansky (Bodansky and Bodansky, 1984) as follows: the amino acid 89 (400 mg, 1 mmol) was dissolved in methanol/water and neutralised with Cs₂CO₃ (300 mg), then the solvents were removed in vacuo, then DMF added and removed in vacuo. The residue was dissolved in DMF (10 mL) and benzyl bromide (190 mg, 1.1 mmol, purified by passage through a short column of basic alumina) added to the stirred solution. After 2 h the reaction was diluted with aq. NaHCO₃ and extracted with 1:1 EtOAc:light pet. The organic layer was washed in turn with aq.NaHCO3, water (x2), brine and then dried over MgSO4. Evaporation of the solvent left the product 90 as a clear oil which solidified to a low melting solid (m.p. ~55°C) on standing (500 mg, TLC 25%EtOAc in light pet. Rf=0.57. 1H NMR (300 MHz, CDCl₃):

7.38-7.32, 4H, m; 7.28-7.14, 6H, m; 5.82, 1H, m; 5.19-5.05, 4H, m's, (OBn ABq, J=12.5 Hz, \Box_a =5.16, \Box_b =5.12 ppm); 4.9, 1H, br; 3.88, 1H, br; 3.44, 1H, bt, J=7 Hz; 2.88, 1H, dd, J=5, 14 Hz; 2.77-2.60, 2H, bm; 1.63, 1H, m; 1.56-1.35, m, 2H; 1.33, 9H, bs (Boc); 0.88, 3H, d, J=6.5 Hz; 0.85, 3H, d, J=6.5 Hz. 13 C NMR (75 MHz, CDCl₃): \Box 175.5; 155.5; 138.6; 135.8; 135.2; 129.2; 128.5; 128.2; 126.1; 117.4; 78.90; 66.40; 58.96; 58.49; 54.25; 42.83; 36.33; 35.71; 28.27 (Boc); 24.77; 22.63; 22.32. Mass Spectrum (ISMS) m/z 495 (M+H+), calculated for C₃₀H₄₂N₂O₄: 494.

The amine **90** (500 mg, 1 mmol) was dissolved in ethylacetate (20 mL) and 37% aqueous formaldehyde solution (0.5 mL) was added. The solution was stirred for 12 h and then diluted with light

10

15

20

25

30

petroleum (40 mL) and washed in turn with aq. NaHCO3, water (x2) and brine and then dried (MgSO₄). Removal of the solvent in vacuo gave the product 91 as a clear oil in approximately quantitative yield. Further purification was carried out by flash chromatography eluting with 10% ethyl acetate in light pet. ¹H NMR (500 MHz, CD₃CN); $\Box\Box$ (rotamers were present in a ratio of 7:3) 7.36, 4H,m, Ar; 7.27-7.11, 6H, Ar; 5.70. 1H, m, vinyl CH; 5.17-4.97, 4H, m's, vinyl CH₂ and OCH₂Ph; 4.44, 0.7H, d, J=5.0 Hz, ring CH₂(a), major rotamer; 4.33, 0.3H, d, J=4.4 Hz, ring CH₂(a), minor rotamer; 4.19, 0.7H, d, J=5.0 Hz, ring CH₂(b), major rotamer; 4.09, 0.3H, d, J=4.6 Hz, ring CH₂(b), minor rotamer; 4.06, 0.3H. m, Phe ; minor; 4.02, 0.7H, m, Phe ; major; 3.74, 0.7H, dd, J=9.8, 6.0 Hz. and 3.69, 0.3H, m, Leu□; 3.10, 1H, m, ring methine (homoallyi); 2.88, 0.3H, m, Phe□(a); 2.84, 0.7H, dd, J=4.1, 13.4, Phe□(a); 2.72. 0.3H, dd, J=6.5, 13.5, Phe□(b); 2.65, 0.7H, dd, J=9.5, 13.2, Phe□(b); 2.49, 1H, m, allyl(a); 2.15, 1H, m, allyl(b); 1.76-1.42, 3H, m's, Leu□+□; 1.33, 2.5H, s, Boc, minor rotamer; 1.09, 6.5H, s, Boc, major rotamer; 0.97-0.84, 6H, d's, Leu□ (major rotamer: 0.94, J=6.3 Hz; 0.90, J=6.2 Hz). 13C NMR (75 MHz, CD₃CN), only major rotamer reported except where indicated: (ref. 118.69 ppm) 173.3; 154.2; 140.9; 137.8; 136.3 (CH); 131.3; 129.9; 129.7; 129.6; 129.5; 127.2; 118.2 (CH₂); 79.98 (Boc tertiary); 67.17 (CH₂); 63.49 (CH); 62.47 (CH₂); 60.91 (CH); 57.68 33.18 (CH₂); (29.08 Boc minor (CH): 40.34 (CH₂); 36.04 (CH₂); rotamer); 28.61 (Boc major rotamer); 25.98 (CH); 23.79 (CH₃); 22.36 Mass Spectrum (ISMS) m/z 507 (MH+), calculated for (CH₃).C₃₁H₄₂N₂O₄: 506.

The alkene 91 was dihydroxylated with OsO_4/N_1 -methylmorpholine-N-oxide in tBuOH/water as previously described for the dihydroxylation of **60**. The crude product **92** was used directly in the next reaction. TLC 1:1 EtOAc:light pet. Rf=0.36. Mass Spectrum (ISMS) m/z 541 (M+H⁺), calculated for $C_{31}H_{44}N_2O_6$: 540.

The glycol 92 (87 mg, 0.16 mmol) was dissolved in THF (4 mL) and H_5IO_6 (37 mg, 0.16 mmol) dissolved in THF (3 mL) was added.

10

15

and the reaction stirred at room temperature. A precipitate of iodic acid rapidly formed and the reaction was complete in <5 min. The THF solution was diluted with ether and washed in turn with 10% aq.Na₂CO₃, water, brine and then dried (MgSO₄). The product aldehyde 93 was of good purity but was not particularly stable to storage. Any traces of acid must be rigorously excluded to prevent isomerisation to the trans isomer. A portion was purified by flash chromatography, eluting with 15%EtOAc in light petroleum. TLC 15%EtOAc in light pet. Rf=0.27. The yield was good (>80%). Amide rotamers were evident in the NMR spectra, ratio ~3:1, only the peak due to the main rotamer is reported unless otherwise noted. 1H NMR (300 MHz, CD₃CN, ref 1.94 ppm): 0 9.53, 1H, s; 7.42-7.10, 10H, m's; 5.11, 2H, s, (OCH₂Ph); 4.41, 1H, br; 4.25, 1H, q, J=6.3 Hz; 4.15, 1H, br; 3.56, 1H, dt, J=8.5, 5.7 Hz; 3.54, 1H, bm; 2.90-2.58, 4H, m; 1.75-1.45, 3H, bm; 1.37, bs, Boc minor rotamer; 1.20, bs, Bocmajor rotamer; 0.92, 3H, d, J=6 Hz; 0.88, 3H, d, J=5.7 Hz. ¹³C NMR (75 MHz, CD₃CN, ref 118.69 ppm):

202.0; 173.1; 154.2; 140.4; 137.6; 131.1; 129.9; 129.62; 129.55; 127.26; 80.28 (Boc tertiary); 67.31 (CH₂); 61.90 (CH₂); 60.43 (CH); 58.56 (CH); 57.95 (CH); 43.75 (CH₂); 40.36 (CH₂); 36.48 (CH₂); 28.66 (Boc); 25.83 (CH); 23.67 (CH₃); 22.25 Mass Spectrum (ISMS) m/z 509 (MH+), calculated for 20 (CH₃). C₃₀H₄₀N₂O₅: 508.

Conversion of 4,5-cis aldehyde 93 to the 4,5-cis 4(S) amine product was completed by a two step reductive amination procedure as illustrated below:

25

10

15

20

25

30

Alanine methyl ester hydrochloride (120 mg, 0.86 mmol) was dissolved in 1:1 brine:10%aq.Na₂CO₃ and extraction into CH₂Cl₂ (x2). The organic extracts were dried (MgSO₄), filtered and the majority of the solvent removed in vacuo to leave the volatile amine which was added to a solution of the freshly prepared aldehyde 93 (100 mg, 0.2 mmol) dissolved in methanol (~7 mL, strictly acid free). The solution was stirred at room temperature for 2 h whereupon analysis of a test portion reduced with NaBH₄ showed imine formation to be complete (none of the alcohol formed on reduction of aldehyde was detected). Solid NaBH₄ (50 mg, 1.3 mmol) was added to the solution and stirring continued for 10 min and between ethyl acetate and reaction partitioned then water/brine/10%aq.Na₂CO₃ mixture. The aqueous phase was separated and the organic layer washed with water (x2) then brine and then dried NMR analysis of the crude product failed to detect the (MgSO₄). corresponding trans (S) diastereomer (<5%). Evaporation of the solvent left an oil which was purified by flash chromatography eluting with 20-40% EtOAc in light petroleum for a 60-70% yield of 94. TLC 40%EtOAc:light Rotamers observed in the NMR spectra, ratio ~3:1, pet. Rf=0.43. separate signals due to the minor rotamer recorded only where indicated. 1H NMR (300 MHz, CD₃CN, ref. 1.94 ppm): □ 7.37, 4H, m,; 7.3-7.1, 6H, m; 5.12, 5.09: 2H, ABq, J=12 Hz; 4.39 (major rotamer), 4.29 (minor): 1H, d. J=5 Hz; 4.15, 1H, J=5 Hz; 4.06, 1H, m, PheHa; 3.75-3.57, 4H, m, LeuH□+OCH₃; 3.25-3.10, 1H, m; 3.03, 1H, m; 2.87-2.60, 2H, m, Phe□; 2.52-2.25, 2H, m; 1.81, 1H, m; 1.67, 1H, m; 1.6-1.38, 2H, m; 1.34, bs, Boc minor rotamer; 1.19, m, Ala ; 1.15, bs, Boc major rotamer; 0.93, 3H, d, J=6.6 Hz; 0.89, 3H, d, J=6.3 Hz. ^{13}C NMR (75 MHz, CD₃CN, ref. 118.69 ppm):

177.3; 173.4; 154.2; 141.0; 137.7; 131.2; 130.9; 129.9; 129.7; 129.6; 129.5; 127.1; 80.02 (Boc tertiary); 67.18 (CH₂); 62.55 (CH); 62.25 (CH₂); 60.75 (CH); 57.67 (2xCH, coincident signals); 52.55 (OCH₃); 45.96 (CH₂); 40.96 (CH₂); 36.15 (CH₂); 29.00 (Boc, minor rotamer); 28.73 (CH₂); 28.62 (Boc, major rotamer); 25.96 (CH);

10

15

20

25

23.66 (CH₃); 22.35 (CH₃); 19.7 (CH₃). Mass Spectrum (ISMS) m/z 596 (M+H $^+$), calculated for C₃₄H₅₀N₃O₆: 595.

Reductive amination of aldehyde **93** (or the 4,5-trans isomer) with NaBH(OAc)₃ in dichloroethane gave rise to a mixture of products **94** and **95** in the ratio 1:9.

The aldehyde 93 (50 mg, 0.1 mmol) was dissolved in 1,2dichloroethane (5 mL) and alanine methyl ester (~2 equivalents) and acetic acid (1drop, ~14 mg) were added. The mixture was stirred at room temperature for 5 min and then NaBH(OAc)3 (40 mg, 2 eq.) was added and stirring continued for 30 min. The solvent was then removed in vacuo and the residue partitioned between EtOAc and 10% aq. Na₂CO₃, the organic layer was washed with water and brine and then dried (MgSO₄). The product contained both diastereomers in the ratio ~9:1, trans:cis. The products were purified by flash chromatography eluting with 20-45% EtOAc in light petroleum. TLC 40% EtOAc:light pet. Rf=0.43 (minor diastereomer, 94, cis), 0.23 (major diastereomer, 95, trans). Combined Rotamers were not observed although significant peak vield ~60%. broadening was present, as observed for the corresponding trans aldehyde. The configuration of the major product was determined by NMR (NOESY experiment). 1H NMR (300 MHz, CD₃CN, ref 1.94 ppm): [] 4.38, 1H, br, ring 5.13, 2H, s, OCH₂Ph; 7.24-7.14, 10H, m's; methylene(i); 3.97, 1H, bd, ring methylene(ii); 3.61, 3H, s. OCH₃; 3.75, 1H, ddd, J=2.7, 4.3, 8.7 Hz, PheH□; 3.50, 1H, m, LeuH□; 3.13, 1H, m_e. PheC'H(ring); 2.97-2.88, 2H, m, AlaH□+PheH□(i); 2.72, 1H, dd, J=2.9, 8.7 Hz, PheH□(ii); 2.33, 1H, ddd, J=11.5, 7.3, 5.5 Hz, CH₂NH(bridge)(i);

10

1.98, 1H, m (dt, overlaps with solvent peak), $CH_2NH(bridge)(ii)$; 1.53, 2H, m, $Leu \square + \square$; 1.43, 9H(s)+1H(m), $Boc+Leu \square$; 1.35, 1H, m, $bridge CH_2(i)$; 1.29, 1H, m, $bridge CH_2(ii)$; 1.06, 3H, d, J=7.0 Hz, $Ala \square$; 0.88, 6H, m, $Leu \square$. ¹³C NMR (75 MHz, CD_3CN , ref 118.69 ppm): \square 177.2; 174.6; 154.5; 140.1; 137.6; 131.0; 129.9; 129.7; 129.6; 127.6; 80.61 (Boctertiary); 67.50 (CH_2); 63.62 (CH_2); 63.5 (CH_1 , br); 62.4 (CH_2 , v.br); 60.67 (CH_2); 57.70 (CH_2); 52.47 (CH_2); 45.15 (CH_2); 40.65 (CH_2 , v.br); 39.76 (CH_2); 32.81 (CH_2); 29.00 (CH_3 , CH_2); 26.21 (CH_2); 23.47 (CH_3); 22.88 (CH_3); 19.62 (CH_3). Mass Spectrum (ISMS) m/z 596 (CH_2), calculated for $C_{34}H_{49}N_3O_6$: 595. ²

The diastereomeric amines were converted to the protected unturn mimetic compounds 96 and 97 as described below:

15

20

The 4,5-cis amine 94 (42 mg, 0.07 mmol) was dissolved in ethyl acetate:ethanol 10:3 (13 mL) and 35 mg of 10% palladium on activated carbon was added and the mixture hydrogenated at 32 psi H₂ for 3 h to deprotect the benzyl ester to the amino acid (MH⁺ = 506 Da). The solution was filtered and the solvent removed *in vacuo*, then the residue was dissolved in DMF (2 mL) and diluted with CH₂Cl₂ (15 mL) and DIEA (50 mg, ~0.4 mmol) and BOP reagent (50 mg, 0.11 mmol) were added to the stirred solution at room temperature. The cyclisation was complete within a few minutes; the CH₂Cl₂ was then removed *in vacuo*

المراعدية

5

10

15

20

25

30

and the residue diluted with ethyl acetate and washed in turn with 10% aq.Na2CO3/brine, water (x2), brine and then dried (MgSO4) and the solvent removed in vacuo to leave a clear oil which was purified by flash chromatography eluting with 20% EtOAc in light petroleum for a yield of 25 mg (70%) of 96. TLC 1:1 EtOAc:light pet ~0.45 The NMR spectra in CD₃CN at room temperature were significantly broadened indicating a degree of conformational interconversion slow on the NMR timescale. ¹H NMR (300 MHz, CD₃CN):

7.32-7.15, 5H, m, Ar; 4.88, 1H, q, J=7.1 Hz, Ala□; 4.20, 1H, bd, J=4.8 Hz, NCH₂N(a); 4.13, 1H, m, Phe□; 4.09, 1H, bd, J=5.0 Hz, $NCH_2N(b)$; 3.72, $\overline{1}H$, m, Leu \Box ; 3.65, 3H, s, OCH_3 ; 3.52, 1H, bdd, J=10.6, 15.2 Hz, bridge CH₂CH₂N(a); 3.30-3.21, 2H, m's CH₂CH₂N(b) and PheC'H; 2.94, 1H, bm, Phe□(a); 2.76. 1H. bm. 2.25 water peak; 1.9-1.4, 5H, e, Leu□+□ and bridge Phe□(b); CH₂CH₂N; 1.29, 3H, d, J=7.1 Hz, Ala ; 3.25, 9H, vbr, Boc; 0.92, 6H, d, J=6.2 Hz, Leu□. ¹³C NMR (75 MHz, CD₃CN): □ 173.5 (the amide and ester peaks appear to be co-incident); 154.9 (carbamate, br); 140.7; 130.7 (br); 129.6; 127.3; 80.54; 66.47; 63.83 (br); 62.36; 60.4 (very br); 56.29; 52.97; 44.77; (36.96, 36.40) very br, just resolved; 33.3 (very br); 28.78 (Boc, br); 26.86; 23.90 (br); 22.63; 15.47. Mass spectrum (ISMS) m/z 250.2 (M+H+), calculated for C28H37N3O6: 511 fragments (OR 60): 441, (-tBu); 397, (-Boc).

The synthesis of **97** was as for **96** but using the trans amine **95**. TLC 1:1 EtOAc:light pet. Rf=0.53. The NMR spectra in CD₃CN were were well resolved and rotamers were present in the ratio of 11:9; signals attributable to the same atom in the different rotamers are placed in parentheses where possible. ¹H NMR (300 MHz, CD₃CN, ref 1.94 ppm): ☐ 7.34-7.16, 5H, m; 4.69, 1H, m; 4.13, 1H, d, J=4.4 Hz; 3.92, 1H, m; (3.83, d, J=4.4; 3.79, d, J=4.4 Hz), 1H; 3.76-3.60, 2H, m's; (3.61, s; 3.81, s), 3H, OCH₃; 3.26, 1H, m; 3.15, 1H, m; 2.99, 1H, m; 2.77, 1H, m; 1.85-1.49, 3H, m's; (1.44, s; 1.41, s), 9H, Boc; 1.30, 3H, d, J=7.2 Hz, Ala□; 1.36-1.24, 2H, m; 0.98-0.91, 6H, m. ¹³C NMR (75 MHz, CD₃CN, ref 118.69 ppm): ☐ 174.4; 173.3; 154.6; (140.54, 140.49); 130.7;

130.6; 129.8; 127.6; (80.65, 80.54), Boc tertiary; (66.12, 65.48, 65.21, 64.90) 2xCH; 60.67, CH₂; (56.82, 56.74), CH; (56.41, 56.24), CH; 52.87, CH₃; (46.19, 46.12), CH₂; (40.72, 39.84), CH₂; 39.16, CH₂; 30.44, CH₂; (29.03, 28.93) Boc; (25.64, 25.58), CH; 24.19, CH₃; 22.43, CH₃; 15.76, CH₃. Mass Spectrum (ISMS) m/z 488 (MH $^+$), calculated for C₂₈H₃₇N₃O₆: 487.

Example (G). Acid catalysed isomerisation of aldehydes 93

10

15

20

25

The trans (4(S)) aldehyde was obtained by the acid catalysed isomerisation of the cis diastereomer 93 in chloroform solution Significant decomposition to multiple. with catalytic HCl present. unidentified by-products (most having high Rf) also occurs under the The product was purified by flash isomerisation conditions. chromatography eluting with 15% ethyl acetate in petroleum ether for a yield of about 35% 98 from crude 93. 1H NMR (300 MHz, CD₃CN, ref. 1.94 ppm):

9.41, t, J=1.8 Hz; 7.45-7.10, 10H, m; 5.12, 2H, m, OCH₂Ph; 4.46, 1H, br; 4.01, 1H, bd; 3.82, 1H, m; 3.62-3.46, 2H, m; 2.95, 1H, bdd, J=13.0, 4.4 Hz; 2.81, 1H, dd, J=13.2, 8.0 Hz; 2.37, 2H, m (ABq of dd, J_{AB} =31, J_{ddA} =4.6, 1.8 Hz; J_{ddB} =7.2, 2.1 Hz), \square -aldehyde; 1.75-1.25, 12H, e (1.4, bs, Boc); 0.9, 6H, bm. ¹³C NMR (75 MHz, CDCl₃):

□ 202.9; 174.5; 154.4; 139.7; 137.5; 131.0; 129.9; 129.8; 129.7; 127.7; 80.81; 67.61; 64.03 (br); 63.18 60.49; 59.9 (br); 47.0 (br); 45.95; 39.88; 28.96 (Boc); 26.12; 23.25; 22.97.

Example (H). Synthesis of a -turn mimetic II(i)

10

15

Compound 70 was prepared as described above, and reacted with alanine methyl ester to form 99 using the same method previously described for the synthesis of 71. The crude amino ketone 99 (1.22g) was reacted with Cbz-glycine symmetric anhydride (synthesised from 1.95g CbzGlyOH and 9.3mls 0.5M dicyclohexylcarbodiimide in dichloromethane) and 0.6g DIEA in dichloromethane. The reaction was stirred at room temperature for 10 hours then diluted with ether (any DCU precipitate was filtered off) and the ether solution was washed with 1M HCI, aqueous sodium bicarbonate then brine and then dried over magnesium sulfate (removed by filtration) and the volatiles removed under reduced pressure to leave the crude product as an oil which was purified by flash chromatography eluting with 2:1 ethyl acetate:light petroleum ether, yield of 100 was 1.8g (90%). Reductive amination of 100 with 101 derived from the deprotection of BocLys(Fmoc)OBn (TFA. CH2Cl2) is carried out by the previously described method for the formation of 73 (71% yield after flash chromatography eluting with 2:1-to

10

15

20

25

30

3:1 ethyl acetate:light petroleum). The product amine 102 was dissolved in ethyl acetate and formalin added to the stirred solution resulting in the formation of imidazolidine 103. The ethyl acetate solution was washed with aqueous sodium bicarbonate, water (twice), brine and then dried over magnesium sulfate (removed by filtration) and the volatiles removed under reduced pressure to leave the crude product as an oil which was purified by flash chromatography eluting with 3:2 ethyl acetate:light petroleum ether (yield >75%). The protected pre-cyclisation compound 103 (400 mgs) was dissolved in 0.1M ethanolic HCI (20 mls) and hydrogenated with 250mgs of 10% Pd-C. The hydrogenation was complete after 7 hours (about 40 psi H₂, room temperature). The solution was filtered through a celite pad to remove the catalyst and 50 mls of DMF added. Volatiles (ethanol) were removed under reduced pressure then a solution of BOP reagent (300 mgs) and DIEA (300 mgs) in 150 mls of DMF was added and the mixture stirred at room temperature for 15 minutes. Most of the DMF was removed under reduced pressure and the residue dissolved in ethyl acetate and washed with 1M HCl, aqueous sodium bicarbonate, water (twice), brine and then dried over magnesium sulfate (removed by filtration) and the volatiles removed under reduced pressure to leave about 300 mgs of crude product 104. product was dissolved in 30 mls methanolic HCI (0.1M) and hydrogenated (200mgs Pd-C, 40psi H₂) for 24 hours reducing the imidazolidine to an N-The catalyst was filtered off (celite) and the solvent methyl group. removed under reduced pressure, the residue was then treated with tetrabutylammonium fluoride in THF to remove the FMOC group. The free amine was then reprotected by addition of benzyl chloroformate (65 mgs) and DIEA (100 mgs). After stirring for 1 hour ethyl acetate was added and the organic layer was washed with 1M HCl, water, then brine, dried over magnesium sulfate (removed by filtration) and the volatiles removed under reduced pressure to leave an oil which was purified by flash chromatography eluting with 3-5% ethanol in chloroform for a yield of about 40% of 105 based on 103.